

### Diagnostic and therapeutic roles of iron oxide nanoparticles in biomedicine

Chia-Hung Lu<sup>a</sup>, Jong-Kai Hsiao<sup>a,b</sup>\*

<sup>a</sup>Department of Medical Imaging, Taipei Tzu Chi Hospital, Buddhist Tzu Chi Medical Foundation, New Taipei, Taiwan, <sup>b</sup>School of Medicine, Tzu Chi University, Hualien, Taiwan

Submission	: 23-Mar-2022
Revision	:03-May-2022
Acceptance	:08-Jun-2022
Web Publication	: 27-Jul-2022

#### INTRODUCTION

#### Nanotechnology in biomedicine

anotechnology is defined as the synthesis and manipulation of materials between 1 and 100 nm [1]. Due to the proximity diameter of iron oxide nanoparticles (IONs) to atoms, these materials exhibit different physical and chemical properties than their corresponding bulk materials. For instance, as the diameter of gold is <30 nm, the color of these gold particles appears ruby red [2]. Furthermore, the surface area of nanomaterials is dramatically increased compared to its volume, making the chemical catalytic process quick. The abundance of the surface area is advantageous when the primary nanoparticles are conjugated with a small ligand, making the particle multifunctional. Nanotechnology-based liposomal drugs have been launched for anticancer efficacy. With different pharmacodynamic/pharmacokinetics of these liposomes compared to their small-molecule form, these liposomes have different anticancer indications [3]. The most famous one is Doxil® (Baxter International, USA), which is the liposomal form of doxorubicin. Unlike doxorubicin which has dose-related cardiotoxicity and myelosuppression, Doxil®

Access this article online					
Quick Response Code:	Website: www.tcmjmed.com				
	DOI: 10.4103/tcmj.tcmj_65_22				

#### Abstract

Nanotechnology changed our understanding of physics and chemics and influenced the biomedical field. Iron oxide nanoparticles (IONs) are one of the first emerging biomedical applications of nanotechnology. The IONs are composed of iron oxide core exhibiting magnetism and coated with biocompatible molecules. The small size, strong magnetism, and biocompatibility of IONs facilitate the application of IONs in the medical imaging field. We listed several clinical available IONs including Resovist (Bayer Schering Pharma, Berlin, Germany) and Feridex intravenous (I.V.)/Endorem as magnetic resonance (MR) contrast agents for liver tumor detection. We also illustrated GastroMARK as a gastrointestinal contrast agent for MR imaging. Recently, IONs named Feraheme for treating iron-deficiency anemia have been approved by the Food and Drug Administration. Moreover, tumor ablation by IONs named NanoTherm has also been discussed. In addition to the clinical application, several potential biomedical applications of IONs including cancer-targeting capability by conjugating IONs with cancer-specific ligands, cell trafficking tools, or tumor ablation agents have also been discussed. With the growing awareness of nanotechnology, further application of IONs is still on the horizon that would shed light on biomedicine.

**Keywords:** Iron oxide, Magnetic resonance imaging, Molecular imaging, Nanotechnology

has higher drug concentration in certain malignancies such as Kaposi's sarcoma and ovarian cancer [4,5].

IONs are one of the nanocomposites that enter the clinical field. Unlike liposomal drugs with small, active molecular components, IONs are composed of an iron oxide core with a hydrophilic coating. The paramagnetic iron oxide core plays an essential part in biomedical applications such as contrast medium for magnetic resonance imaging (MRI) or magnetic beads for specific cell sorting [6]. The scale comparison of IONs, liposomes, and the scale of natural materials is illustrated in Figure 1.

## Physics and biochemical properties of iron oxide nanoparticles

IONs are magnetite (FeO'Fe<sub>2</sub>O<sub>3</sub>) or maghemite ( $\gamma$ -Fe<sub>2</sub>O<sub>3</sub>) core [7,8]. The core is not water soluble. Therefore, coating with water-soluble polymers such as dextran, carboxydextran,

For reprints contact: WKHLRPMedknow\_reprints@wolterskluwer.com

How to cite this article: Lu CH, Hsiao JK. Diagnostic and therapeutic roles of iron oxide nanoparticles in biomedicine. Tzu Chi Med J 2023;35(1):11-7.

<sup>\*</sup>Address for correspondence: Dr. Jong-Kia Hsiao, Department of Medical Imaging, Taipei Tzu Chi Hospital, Buddhist Tzu Chi Medical Foundation, 289, Jianguo Road, Xindian District, New Taipei, Taiwan. E-mail: jongkai@gmail.com

This is an open access journal, and articles are distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 4.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as appropriate credit is given and the new creations are licensed under the identical terms.



Figure 1: Scale comparison of Biomedical Nanomaterials and Natural Materials

or polyethylene glycol (PEG) is mandatory. A small molecule such as citrate or glutathione coating for the synthesis has also been tried [9,10]. Magnetite is the most magnetic natural mineral [11]. However, for the synthesis of nanosized iron oxide core, the bottom-up chemical synthesis methods are preferred over top-down physical methods [12]. Some biological techniques such as bacterial synthesis of IONs have also been tried [13]. The size of the iron oxide core is heavily dependent on the synthesis methods, which could also influence the magnetic properties. The selection of coating of iron oxide core is dependent on the purpose of the IONs. However, water-solubility, biocompatibility, and less immunogenicity are three of the necessities [14]. For most of the current clinical available I.V. MR contrast agent, IONs are coated with dextran (Feridex I.V./Endorem) or carboxydextran (Resovist), which could be metabolized by liver and kidneys. Attempts to coat iron oxide core with water-soluble silicates have been tried with successful clinical results such as GastroMARK [15,16]. Although silicates cannot be metabolized by liver and kidneys, oral ingestion is a safe delivery method. These silicate-coated IONs are currently available for gastrointestinal MR contrast medium, which could pass through gastrointestinal tract without retention in the body.

All of the IONs have magnetism. Under 10 nm, the iron oxide core is paramagnetic, which means that the magnetic property of the iron oxide core could reach the saturated magnetism exposed to a high magnetic field [17]. However, the magnetism of the iron oxide core could be reversed once if the polarity of the magnetic field has been reversed. Most importantly, the magnetism of the iron oxide core could return to zero when the surrounding magnetic field was removed [18]. The assurance of the paramagnetism of the iron oxide core could avoid hazardous clustering of IONs after exposure to a high magnetic field such as MRI [14]. According to the hysteresis loop study, Resovist has the coercive field (Hc) of zero and saturated magnetism of 80 emu/g [19].

#### Metabolization of iron oxide nanoparticles

The biological reaction and metabolization of IONs is dependent on the size, surface charge, and coating of the IONs. Consequently, these IONs are mostly located at the liver, where RES is abundant. For those IONs diameter <30 nm, the lymphatic system will recognize these IONs, and most of these IONs are accumulated in the lymph nodes or spleen. If the IONs are designed not to be recognized by immune cells, PEG coating is mostly used [20,21]. Strategies of PEG binding antibody drug are often used in the pharmaceutical

industry to prolong the antibody drug circulation time in the blood stream [22]. The PEG conjugation of the IONs is significant if targeted molecular imaging is required [23]. Once RES takes up the IONs, the surface coating of the carbohydrates will be metabolized, whereas the iron oxide core is transformed into an iron storage pool to synthesize the hemoglobin [21]. Consequently, the IONs with either PEG or dextran/carboxydextran coating are completely biocompatible and could be metabolized as iron source or eliminated to bile or urine. As to the silicate-coating IONs, the delivery route is oral and could be passed into gastrointestinal tract without biological interaction with the human body. These silicate-coating IONs are eliminated as feces.

## Magnetic resonance imaging and iron oxide nanoparticles

The most important clinical application of IONs is based on MRI. The IONs exhibit both T1 and T2 effects. MR T1 effect refers to the reduction of water molecule movement frequency to the resonance state, in which the regional tissue will exhibit higher signal intensity. The MR T2 effect refers to the disruption of the magnetic field in the microenvironment. with sequential reduction of the signal intensity of adjacent tissue. The quantitative expression of the MR T1 and T2 effect is the R1 and R2 value, which is defined by the capability of reduction of the T1 or T2 relaxation time [24]. For most of the gadolinium-based contrast agents, their contrast capability is based on MR T1 effect. However, we can observe both T1 and T2 effect of the IONs that is dependent on the concentration of the IONs and the iron oxide core size. Resovist has a T1 effect in the arterial phase, in which IONs are limited in the great arteries. The MR T2 effect refers to the disruption of the magnetic field in the microenvironment. The R1 and R2 of Resovist is 72 mM<sup>-1</sup> S<sup>-1</sup> and 8.2 mM<sup>-1</sup>·S<sup>-1</sup>, respectively [25]. In comparison with gadolinium-based MR contrast medium such as Omniscan that exhibiting R1 and R2 of 4.9 mM<sup>-1</sup> S<sup>-1</sup> and 4.8 mM<sup>-1</sup> S<sup>-1</sup>, the IONs has a stronger T2 relaxation effect [26]. The IONs are regarded as small magnets capable of interfering with the high magnetic field of the MRI, allowing the contrast capability to existing. Most of the IONs is taken advantage of by the T2 effect [27].

Consequently, T2-weighted sequences are preferred in the applications of IONs [28]. Recently, MR pulse sequences and magnetic particle imaging, designed specifically to IONs, have been developed [29]. The pulse sequence has higher sensitivity to the IONs and better temporal resolution with sparing the exact spatial resolution, making it the best pulse sequence for the IONs [29].

#### Cellular uptake of iron oxide nanoparticles

The cellular uptake mechanism varies according to the size, surface charge, and coating materials of the IONs [30]. The uptake of Resovist by macrophage (Raw264.7, THP-1) and lung cancer cell line (A549) and breast cancer cell line (MDA) is through clathrin receptor-mediated endocytosis [25,31]. The ferumoxide uptake mechanism is also through clathrin receptor-mediated endocytosis evidenced by human promonocyte (THP-1) [32]. Transmission electron microscopy of the cells ingested IONs showed iron oxide core in the membrane-bound vesicles, and lysosomes are believed to be the organelles responsible for the metabolism of IONs [33].

#### Iron oxide nanoparticle on cell proliferation

In the diagnostic dose, the clinical IONs such as Resovist do not influence macrophage nor human mesenchymal cell proliferation [33,34]. Similar findings have been discovered in other clinical IONs such as Endorem® on muscle precursor cells or Feridex® on the bone marrow-derived stem-progenitor cells [35,36]. Different clinically available IONs have also been verified on the same platform, showing the inert property toward human mesenchymal stem cells (MSCs) [37]. Although there is no cellular viability change in the recommended dose, several in vitro studies showed enhanced cell proliferation on the Resovist-labeled cells, including human mesenchymal cells or mouse macrophage [33,34,38]. Cell proliferation is promoted through the depletion of intracellular H<sub>2</sub>O<sub>2</sub>, which further influences the cell cycle. These properties might have applications in tissue engineering, in which cell proliferation is anticipated.

Due to the uptake of IONs by the RES in humans, lots of attention have been paid to the RES-related cellular response after ingestion of IONs. Resovist has been evaluated toward murine macrophage (Raw264.7) cell lines or mouse peritoneal macrophages [34,39]. No cytokine release is found at the recommended dosage in these studies. However, elevated tissue necrosis factor (TNF)- $\alpha$  and interleukin (IL) 1- $\beta$ , as well as IL-6, have been found after the cells were exposed to 100 ug Fe/mL for 24 h [34,39]. Nitric oxide secretion was also elevated under these conditions. Ferumoxides exposed to mouse peritoneal macrophage at a 100 ug Fe/mL concentration for 48 h also have a mild elevation of IL-1 after lipopolysaccharide stimulation [32]. Considering the rapid half-life of IONs in the human body and the clinically suggested dosage, cytokine release after ION injection could be neglected.

#### Iron oxide nanoparticles on stem cell differentiation

In addition to liver imaging, IONs have drawn plenty of attention to stem cell trafficking. The confirmation of inner properties toward cell differentiation is one of the major concerns. Studies on the Resovist toward human MSCs for bone, cartilage, and adipocyte differentiation show no differentiation capacity alteration at 100 ug Fe/mL concentration for 24 h [33]. Interestingly, under such ION conditions, human MSCs could also transdifferentiated into neuron-like cells, with electrical spiking activity being found on these ION-labeled cells [40]. The transdifferentiation of human MSCs into dopaminergic neurons after Resovist labeling is also found [41]. Feridex<sup>®</sup> along with protamine sulfate transfection shows preserved differentiation capacity on hematopoietic or MSCs [42]. These findings support using IONs as a stem cell trafficking tool under MRI.

# CLINICAL APPLICATION OF IRON OXIDE NANOPARTICLES

Inspired by the promising *in vitro* and *in vivo* studies, several clinical trials on IONs passed the American Food

and Drug Administration (FDA), European Medicines Agency (EMA), and Taiwan Food and Drug Administration. We listed IONs launched in the clinical fields and listed some IONs under clinical trials in Table 1.

Currently, IONs have been applied in medical diagnostic imaging for detecting liver tumor [27]. The most well-known IONs in the clinical fields are Resovist and Feridex I.V. In addition to liver tumor detection, IONs are also used as gastrointestinal negative contrast agents and manipulated as slow-release iron supplements for treating iron-deficiency anemia. These clinically available IONs are described as follows.

#### Resovist

Resovist, a T2 superparamagnetic ION, was primarily designed for liver imaging. The uptake of IONs by RES has a strong susceptibility effect that causes loss of signal intensity on T1- and T2-weighted images, making organs containing RES appear dark, primarily in the liver, spleen, and also in bone marrow and lymph nodes.

Kupffer cells in the liver take up IONs which shorten T1 and T2/T2\* relaxation times. Normal liver parenchyma and lesions containing Kupffer cells include hepatic adenoma, focal nodular hyperplasia (FNH), cirrhotic regenerative nodules, and some well-differentiated nodules hepatocellular carcinomas (HCCs) usually show dark signal intensity on ION-enhanced images [45-47]. In contrast, tumors that do not contain Kupffer cells such as metastases and cholangiocarcinoma show high signal intensity on ION-enhanced images. The use of ION contrast agent in MRI is proved to be a valuable tool for differentiating between HCCs and other benign focal hepatic lesions [48-50]. A meta-analysis showed a sensitivity of 85% and a specificity of 78% [48]. Quantitative measurement of late-phase enhancement after slow Resovist administration also differentiates malignant from benign hepatic masses and hemangiomas from other focal liver lesions [51].

Differentiating HCC from FNH is among the most critical applications. Many FNH has typical image findings, such as the presence of a central scar. In some cases, they may mimic well-differentiated HCC. However, many other FNH may still show atypical findings, making the diagnosis difficult [52]. In these cases, IONs are a valuable tool in differentiating HCC from FNH [53]. This imaging strategy is of great use because a completely different treatment strategy is required for HCC and FNH.

Differentiating HCC from DN is also another important application of ION contrast for early and precise treatment of HCC in cirrhotic liver [45,46]. Histologic observations proposed that HCCs are thought to develop from a regenerative nodule to a dysplastic nodule, subsequently into a well-differentiated HCC and eventually to moderately and poorly differentiated HCC [54]. As the liver's signal intensity changes and rich Kupffer cells are positively correlated, there is an inverse correlation between Kupffer cell count and the degree of HCC differentiation, indicating that the uptake of IONs in HCCs decreased as the degree of differentiation of HCCs declined [55]. Therefore, ION-enhanced MRI may

IONs proved by FDA/EMA/TFDA							
Brand name	Generic name	Size (nm)	Surface coating	Indication	<b>Delivery methods</b>	Clinical status	
Resovist	Ferucarbotran	60	Carboxydextran	Liver tumor detection	I.V. slowly push	EMA/TFDA approval	
Feridex I.V.	Ferumoxides	80-150	Dextran	Liver tumor detection	I.V. drip	FDA approval	
Endorem	Ferumoxides	80-150	Dextran	Liver tumor detection	I.V. drip	EMA approval	
Gastromark	Ferumoxsil	200-400	Silica	GI tract imaging	Oral	FDA approval	
Faraheme	Ferumoxytol	30	Polyglucose sorbitol carboxymethylether	Iron-deficiency anemia	I.V. drip	FDA approval	
NanoTherm	NanoTherm	10-15	Aminosilane	Thermal ablation of GBM	Local injection	EMA approval	
(ONe: Iron oxide nanonarticles, EDA: Food and Drug Administration, EMA: European Medicines Agency, TEDA: Taiwan EDA, GI: Gastrointestinal, GBM:							

Table 1: Iron oxide nanoparticles proved by Food and Drug Administration/European Medicines Agency/Taiwan Food and Drug Administration[43,44]

IONs: Iron oxide nanoparticles, FDA: Food and Drug Administration, EMA: European Medicines Agency, TFDA: Taiwan FDA, GI: Gastrointestinal, GBM: Glioblastoma, I.V.: Intravenous

play a beneficial role in differentiating DN from poorly differentiated HCC.

#### Feridex I.V./Endorem

Feridex I.V. (Berlex Laboratories, USA) are FDA approved IONs coated by dextran with hydroxyl diameters of 80–150 nm. Endorem (Guerbet, Villepinte, France) is the corresponding IONs licensed by EMA. Like Resovist, the Feridex I.V. could be phagocytosed by RES in the liver, allowing good contrast between normal and tumorous liver tissue. One study focusing on the optimizing MR pulse sequence for Feridex shows that the identification of liver tumors could reach 0.3 cm [28]. However, unlike Resovist, which could be delivered by I.V. push, Feridex I.V. could only be administered by I.V. drip. For the cell labeling that is widely used in the cell therapy study, Feridex I.V. is usually mixed with a transfecting agent or protamine sulfate to enhance its labeling efficiency [42].

#### GastroMARK

GastroMARK is IONs coated with silica at the 200–400 nm diameter. Unlike other biocompatible IONs, GastroMARK is orally ingested and eliminated as feces. Under MR scanning, the darkening effect of GastroMARK contrasts the lumen and edematous bowel that aids in the diagnosis of inflammatory bowel disease [16]. Moreover, GastroMARK has been used as a negative contrast agent to reduce intestinal juice that could interfere with the biliary tree and pancreatic duct signals [56,57]. However, patients receiving GastroMARK experienced metallic taste that is unpleasing, which limited its acceptance in the clinical field [57].

#### Feraheme

Feraheme is composed of mixed magnetite/maghemite cores about 3- to 10 nm with carboxymethyl dextran coating that made up the whole particle of 30 nm [58,59]. Unlike most of the clinical application of IONs on MRI, Feraheme has been approved by the FDA for the treatment of iron-deficiency anemia. Traditional I.V. iron supplement such as iron sucrose could only be delivered in a small amount of 100–200 mg, but Feraheme could be administered at a larger amount of 510 mg [60]. The delivery amount is critical especially for chronic renal failure patient, in whom 1000 mg of iron should be given in each course. Feraheme has better dosing advantage and the incidence and severity of adverse reaction is identical to iron sucrose [60]. Owing to the clinical availability and

safety evaluation, more potential applications of Feraheme have been investigated such as treating ferroportin-deficient leukemia or MR hyperthermia agent [61,62].

#### TRANSLATIONAL AND EXPERIMENTAL

#### INVESTIGATIONS OF IRON OXIDE NANOPARTICLES Molecular imaging and therapy

Unlike fluorescent imaging, which can only detect tissues within centimeters, MR has unlimited tissue detection depth, making it an ideal noninvasive diagnostic tool. Moreover, the ionizing radiation-free character makes MR a better imaging modality than computed tomography for screening or follow-up. The iron oxide that is conjugated with cancer-specific targeting molecules could then be used for cancer imaging. For example, one breast cancer-specific peptide that was discovered by phage display was conjugated with IONs for breast cancer imaging. In vitro, ex vivo, and in vivo mouse xenografts show specific binding of these IONs to the cancer cells. Moreover, MR successfully identified these xenografts after ION delivery [63]. This conceptual diagnostic strategy has also been verified in prostate cancer and lung cancer [23,64]. The cancer-specific antibody that was used for cancer therapy, such as Herceptin, was also conjugated to IONs for specific cancer targeting [65-67]. However, these cancer-targeting effects could mostly be visualized in the in vitro study. The probable cause is the relatively large molecular weight of Herceptin compared with cancer-specific peptides, making it hard to penetrate the tumor capillaries. Multiple antibody binding sites to the IONs might also alleviate its potency toward cancer cells. Optimizing the targeting molecular size and the whole diameter of the IONs is still one of the challenging works.

In addition to targeted imaging, IONs exposed to alternating magnetic fields could convert these energies into thermal energy. These thermal effects could facilitate chemotherapy, radiation therapy, or immunotherapy. For example, NanoTherm, an ION with aminosilane coating, has been investigated under clinical trials for treating prostate cancer and got the FDA's approval for ablating glioblastoma multiforme [14,68].

#### Stem cell trafficking

There is great interest in stem cell therapy and trafficking. Once the IONs are labeled into stem cells, they could be traced by MRI, making it an ideal strategy for validating the stem cell fate and location [69,70]. For example, in a mouse glioblastoma model, human MSCs labeled with Resovist were injected into the mouse brain and monitored for the interaction of tumor size and human MSC migration. Under MRI serial following, the glioma xenograft decreased in size, and there is significant human MSC migration toward the glioma [70]. Another example is the ischemic rat model that rat ION-labeled MSCs have injected into ventricular space. The interaction of rat MSCs with choroid plexus is found. Moreover, the ischemic stroke size of this rat MSC-treated group has greater brain recovery [69]. With the growing awareness of cell therapy in regenerative medicine, the pursuit of an ideal imaging method will trigger the development of IONs.

#### Immune cell trafficking

Immune cells such as lymphocytes or monocytes are circulating cells that are hard to trace. However, the accumulation of these cells is positively related to the disease process such as arthritis. Loading immune cells with IONs would be a novel strategy for monitoring disease status. By conjugating PEG-coated IONs with CD3-ab, rat arthritis could be visualized by 4.7T MRI *in vivo* [71]. Vulnerable plaque is the easily rupture plaque that contributes to acute myocardial infarction. Myeloperoxidase is an indicator for vulnerable atherosclerotic plaque. By targeting myeloperoxidase with appropriate ligand, 5-hydroxytryptamine with IONs, MRI successfully demonstrated the aortic atherosclerotic plaque in ApoE-/-mice [72]. These results might shed light on the diagnosis of inflammatory disease.

#### CONCLUSION

With the rapid progress of nanotechnology, growing numbers of IONs are entering clinical trials. The interaction between IONs and the human immune system shaped the biomedical application of IONs. Future targeted molecular imaging is anticipated based on current ION contrast agents. Slow-release iron supplements for treating anemia would be another significant advance in nanotechnology. Tumor ablation by IONs with imaging tumor capability would be other novel advances in tumor theranostics.

#### Financial support and sponsorship

This research was funded in part by the Taiwan Ministry of Science and Technology (MOST-110-2314-B-303-013-MY3, approved in Aug. 2021) and Taipei Tzu Chi Hospital (TCRD-TPE-109-35, approved in Jan, 2019, TCRD-TPE-106-34, approved in Jan. 2017).

#### Conflicts of interest

Dr. Jong-Kai Hsiao, an editorial board member at *Tzu Chi Medical Journal*, had no role in the peer-review process of or decision to publish this article. The other author declared no conflicts of interest in writing this paper.

#### References

- Kaehler T. Nanotechnology: Basic concepts and definitions. Clin Chem 1994;40:1797-9.
- Mulvaney P. Nanoscience versus nanotechnology Defining the field. ACS Nano 2015;9:2215-7.
- 3. Ait-Oudhia S, Mager DE, Straubinger RM. Application of

pharmacokinetic and pharmacodynamic analysis to the development of liposomal formulations for oncology. Pharmaceutics 2014;6:137-74.

- Cagel M, Grotz E, Bernabeu E, Moretton MA, Chiappetta DA. Doxorubicin: Nanotechnological overviews from bench to bedside. Drug Discov Today 2017;22:270-81.
- Norouzi M, Amerian M, Amerian M, Atyabi F. Clinical applications of nanomedicine in cancer therapy. Drug Discov Today 2020;25:107-25.
- Etemadi H, Buchanan JK, Kandile NG, Plieger PG. Iron oxide nanoparticles: Physicochemical characteristics and historical developments to commercialization for potential technological applications. ACS Biomater Sci Eng 2021;7:5432-50.
- Ajinkya N, Yu X, Kaithal P, Luo H, Somani P, Ramakrishna S. Magnetic Iron oxide nanoparticle (IONP) synthesis to applications: Present and future. Materials (Basel) 2020;13:4644.
- Wu W, He Q, Jiang C. Magnetic iron oxide nanoparticles: Synthesis and surface functionalization strategies. Nanoscale Res Lett 2008;3:397-415.
- Kotsmar C, Yoon KY, Yu H, Ryoo SY, Barth J, Shao S, et al. Stable citrate-coated Iron oxide superparamagnetic nanoclusters at high salinity. Ind Eng Chem Res 2010;49:12435-43.
- Liu CL, Peng YK, Chou SW, Tseng WH, Tseng YJ, Chen HC, et al. One-step, room-temperature synthesis of glutathione-capped iron-oxide nanoparticles and their application in *in vivo* T1-weighted magnetic resonance imaging. Small 2014;10:3962-9.
- Harrison RJ, Dunin-Borkowski RE, Putnis A. Direct imaging of nanoscale magnetic interactions in minerals. Proc Natl Acad Sci U S A 2002;99:16556-61.
- Samrot AV, Sahithya CS, Selvarani AJ, Purayil SK, Ponnaiah P. A review on synthesis, characterization and potential biological applications of superparamagnetic Iron oxide nanoparticles. Curr Opin Green Sustain Chem 2021;4:100042.
- Madubuonu N, Aisida SO, Ali A, Ahmad I, Zhao TK, Botha S, et al. Biosynthesis of iron oxide nanoparticles via a composite of psidium guavaja-Moringa oleifera and their antibacterial and photocatalytic study. J Photochem Photobiol B 2019;199:111601.
- Tong S, Zhu H, Bao G. Magnetic Iron oxide nanoparticles for disease detection and therapy. Mater Today (Kidlington) 2019;31:86-99.
- Liu HM, Wu SH, Lu CW, Yao M, Hsiao JK, Hung Y, et al. Mesoporous silica nanoparticles improve magnetic labeling efficiency in human stem cells. Small 2008;4:619-26.
- Gee MS, Harisinghani MG. MRI in patients with inflammatory bowel disease. J Magn Reson Imaging 2011;33:527-34.
- Teja AS, Koh PY. Synthesis, properties, and applications of magnetic Iron oxide nanoparticles. Prog CrystGrowth Charact Mater 2009;55:22-45.
- Hsiao JK, Tai MF, Yang CY, Chen ST, Wang JL, Ku HC, et al. Comparison of micrometer and nanometer sized magnetic particles for cell labeling. IEEE Trans Magn 2007;43:2421-3.
- Yang CY, Tai MF, Chen ST, Wang YT, Chen YF, Hsiao JK, et al. Labeling of human mesenchymal stem cell: Comparison between paramagnetic and superparamagnetic agents. AJP 2009;105:07B314.
- Gómez-Vallejo V, Puigivila M, Plaza-García S, Szczupak B, Piñol R, Murillo JL, et al. PEG-copolymer-coated iron oxide nanoparticles that avoid the reticuloendothelial system and act as kidney MRI contrast agents. Nanoscale 2018;10:14153-64.
- Feng Q, Liu Y, Huang J, Chen K, Huang J, Xiao K. Uptake, distribution, clearance, and toxicity of iron oxide nanoparticles with different sizes and coatings. Sci Rep 2018;8:2082.
- 22. Chapman AP. PEGylated antibodies and antibody fragments for improved therapy: A review. Adv Drug Deliv Rev 2002;54:531-45.
- Yeh CY, Hsiao JK, Wang YP, Lan CH, Wu HC. Peptide-conjugated nanoparticles for targeted imaging and therapy of prostate cancer. Biomaterials 2016;99:1-15.
- Jacques V, Dumas S, Sun WC, Troughton JS, Greenfield MT, Caravan P. High-relaxivity magnetic resonance imaging contrast agents. Part 2.

Optimization of inner- and second-sphere relaxivity. Invest Radiol 2010;45:613-24.

- Yang CY, Tai MF, Lin CP, Lu CW, Wang JL, Hsiao JK, et al. Mechanism of cellular uptake and impact of ferucarbotran on macrophage physiology. PLoS One 2011;6:e25524.
- Dutta S, Kim SK, Lee EJ, Kim TJ, Kang DS, Chang YM, et al. Synthesis and magnetic relaxation properties of paramagnetic Gd-complexes of new DTPA-bis-amides. The X-ray crystal structure of Gd (L)(H<sub>2</sub>O) 3H<sub>2</sub>O (L=DTPA-bis (4-carboxylicphenyl) amide). Bull Korean Chem Soc 2006;27:1038-42.
- Reimer P, Balzer T. Ferucarbotran (Resovist): A new clinically approved RES-specific contrast agent for contrast-enhanced MRI of the liver: Properties, clinical development, and applications. Eur Radiol 2003;13:1266-76.
- Kim SH, Choi D, Lim JH, Lee WJ, Jang HJ, Lim HK, et al. Optimal pulse sequence for ferumoxides-enhanced MR imaging used in the detection of hepatocellular carcinoma: A comparative study using seven pulse sequences. Korean J Radiol 2002;3:87-97.
- Wu LC, Zhang Y, Steinberg G, Qu H, Huang S, Cheng M, et al. A review of magnetic particle imaging and perspectives on neuroimaging. AJNR Am J Neuroradiol 2019;40:206-12.
- Singh N, Jenkins GJ, Asadi R, Doak SH. Potential toxicity of superparamagnetic iron oxide nanoparticles (SPION). Nano Rev 2010;1:5358.
- Guggenheim EJ, Rappoport JZ, Lynch I. Mechanisms for cellular uptake of nanosized clinical MRI contrast agents. Nanotoxicology 2020;14:504-32.
- Raynal I, Prigent P, Peyramaure S, Najid A, Rebuzzi C, Corot C. Macrophage endocytosis of superparamagnetic iron oxide nanoparticles: Mechanisms and comparison of ferumoxides and ferumoxtran-10. Invest Radiol 2004;39:56-63.
- 33. Hsiao JK, Tai MF, Chu HH, Chen ST, Li H, Lai DM, et al. Magnetic nanoparticle labeling of mesenchymal stem cells without transfection agent: Cellular behavior and capability of detection with clinical 1.5 T magnetic resonance at the single cell level. Magn Reson Med 2007;58:717-24.
- Hsiao JK, Chu HH, Wang YH, Lai CW, Chou PT, Hsieh ST, et al. Macrophage physiological function after superparamagnetic iron oxide labeling. NMR Biomed 2008;21:820-9.
- 35. Azzabi F, Rottmar M, Jovaisaite V, Rudin M, Sulser T, Boss A, et al. Viability, differentiation capacity, and detectability of super-paramagnetic iron oxide-labeled muscle precursor cells for magnetic-resonance imaging. Tissue Eng Part C Methods 2015;21:182-91.
- Meng Y, Zhang F, Blair T, Gu H, Feng H, Wang J, et al. MRI of auto-transplantation of bone marrow-derived stem-progenitor cells for potential repair of injured arteries. PLoS One 2012;7:e31137.
- Kim HS, Oh SY, Joo HJ, Son KR, Song IC, Moon WK. The effects of clinically used MRI contrast agents on the biological properties of human mesenchymal stem cells. NMR Biomed 2010;23:514-22.
- Huang DM, Hsiao JK, Chen YC, Chien LY, Yao M, Chen YK, et al. The promotion of human mesenchymal stem cell proliferation by superparamagnetic Iron oxide nanoparticles. Biomaterials 2009;30:3645-51.
- Yeh CH, Hsiao JK, Wang JL, Sheu F. Immunological impact of magnetic nanoparticles (Ferucarbotran) on murine peritoneal macrophages. J Nanoparticle Res 2010;12:151-60.
- Lu CW, Hsiao JK, Liu HM, Wu CH. Characterization of an iron oxide nanoparticle labelling and MRI-based protocol for inducing human mesenchymal stem cells into neural-like cells. Sci Rep 2017;7:3587.
- Chung TH, Hsu SC, Wu SH, Hsiao JK, Lin CP, Yao M, et al. Dextran-coated iron oxide nanoparticle-improved therapeutic effects of human mesenchymal stem cells in a mouse model of Parkinson's disease. Nanoscale 2018;10:2998-3007.
- 42. Wang YX. Superparamagnetic iron oxide based MRI contrast agents:

Current status of clinical application. Quant Imaging Med Surg 2011;1:35-40.

- Anselmo AC, Mitragotri S. A review of clinical translation of inorganic nanoparticles. AAPS J 2015;17:1041-54.
- 44. Arbab AS, Yocum GT, Rad AM, Khakoo AY, Fellowes V, Read EJ, et al. Labeling of cells with ferumoxides-protamine sulfate complexes does not inhibit function or differentiation capacity of hematopoietic or mesenchymal stem cells. NMR Biomed 2005;18:553-9.
- 45. Macarini L, Milillo P, Cascavilla A, Scalzo G, Stoppino L, Vinci R, et al. MR characterisation of dysplastic nodules and hepatocarcinoma in the cirrhotic liver with hepatospecific superparamagnetic contrast agents: Pathological correlation in explanted livers. Radiol Med 2009;114:1267-82.
- Lim JH, Choi D, Cho SK, Kim SH, Lee WJ, Lim HK, et al. Conspicuity of hepatocellular nodular lesions in cirrhotic livers at ferumoxides-enhanced MR imaging: Importance of Kupffer cell number. Radiology 2001;220:669-76.
- Tanaka M, Nakashima O, Wada Y, Kage M, Kojiro M. Pathomorphological study of Kupffer cells in hepatocellular carcinoma and hyperplastic nodular lesions in the liver. Hepatology 1996;24:807-12.
- Li YW, Chen ZG, Wang JC, Zhang ZM. Superparamagnetic iron oxide-enhanced magnetic resonance imaging for focal hepatic lesions: Systematic review and meta-analysis. World J Gastroenterol 2015;21:4334-44.
- 49. Maurea S, Mainenti PP, Tambasco A, Imbriaco M, Mollica C, Laccetti E, et al. Diagnostic accuracy of MR imaging to identify and characterize focal liver lesions: Comparison between gadolinium and superparamagnetic iron oxide contrast media. Quant Imaging Med Surg 2014;4:181-9.
- Bartolotta TV, Midiri M, Galia M, Carcione A, De Maria M, Lagalla R. Benign hepatic tumors: MRI features before and after administration of superparamagnetic contrast media. Radiol Med 2001;101:219-29.
- Santoro L, Grazioli L, Filippone A, Grassedonio E, Belli G, Colagrande S. Resovist enhanced MR imaging of the liver: Does quantitative assessment help in focal lesion classification and characterization? J Magn Reson Imaging 2009;30:1012-20.
- Yoon JH, Kim JY. Atypical findings of focal nodular hyperplasia with gadoxetic acid (Gd-EOB-DTPA)-enhanced magnetic resonance imaging. Iran J Radiol 2014;11:e9269.
- 53. Vogl TJ, Hammerstingl R, Schwarz W, Kümmel S, Müller PK, Balzer T, et al. Magnetic resonance imaging of focal liver lesions. Comparison of the superparamagnetic iron oxide resovist versus gadolinium-DTPA in the same patient. Invest Radiol 1996;31:696-708.
- Sakamoto M, Hirohashi S, Shimosato Y. Early stages of multistep hepatocarcinogenesis: Adenomatous hyperplasia and early hepatocellular carcinoma. Hum Pathol 1991;22:172-8.
- 55. Imai Y, Murakami T, Yoshida S, Nishikawa M, Ohsawa M, Tokunaga K, et al. Superparamagnetic iron oxide-enhanced magnetic resonance images of hepatocellular carcinoma: Correlation with histological grading. Hepatology 2000;32:205-12.
- Frisch A, Walter TC, Hamm B, Denecke T. Efficacy of oral contrast agents for upper gastrointestinal signal suppression in MRCP: A systematic review of the literature. Acta Radiol Open 2017;6:2058460117727315.
- Delaney L, Applegate KE, Karmazyn B, Akisik MF, Jennings SG. MR cholangiopancreatography in children: Feasibility, safety, and initial experience. Pediatr Radiol 2008;38:64-75.
- Wei H, Bruns OT, Kaul MG, Hansen EC, Barch M, Wiśniowska A, et al. Exceedingly small iron oxide nanoparticles as positive MRI contrast agents. Proc Natl Acad Sci U S A 2017;114:2325-30.
- Wang G, Serkova NJ, Groman EV, Scheinman RI, Simberg D. Feraheme (Ferumoxytol) is recognized by proinflammatory and anti-inflammatory macrophages via scavenger receptor type AI/II. Mol Pharm 2019;16:4274-81.

- Macdougall IC, Strauss WE, McLaughlin J, Li Z, Dellanna F, Hertel J. A randomized comparison of ferumoxytol and iron sucrose for treating iron deficiency anemia in patients with CKD. Clin J Am Soc Nephrol 2014;9:705-12.
- Trujillo-Alonso V, Pratt EC, Zong H, Lara-Martinez A, Kaittanis C, Rabie MO, et al. FDA-approved ferumoxytol displays anti-leukaemia efficacy against cells with low ferroportin levels. Nat Nanotechnol 2019;14:616-22.
- Bullivant JP, Zhao S, Willenberg BJ, Kozissnik B, Batich CD, Dobson J. Materials characterization of Feraheme/ferumoxytol and preliminary evaluation of its potential for magnetic fluid hyperthermia. Int J Mol Sci 2013;14:17501-10.
- Hsiao JK, Wu HC, Liu HM, Yu A, Lin CT. A multifunctional peptide for targeted imaging and chemotherapy for nasopharyngeal and breast cancers. Nanomedicine 2015;11:1425-34.
- 64. Chi YH, Hsiao JK, Lin MH, Chang C, Lan CH, Wu HC. Lung cancer-targeting peptides with multi-subtype indication for combinational drug delivery and molecular imaging. Theranostics 2017;7:1612-32.
- Chen TJ, Cheng TH, Chen CY, Hsu SC, Cheng TL, Liu GC, et al. Targeted herceptin-dextran iron oxide nanoparticles for noninvasive imaging of HER2/neu receptors using MRI. J Biol Inorg Chem 2009;14:253-60.
- Mi Y, Liu X, Zhao J, Ding J, Feng SS. Multimodality treatment of cancer with herceptin conjugated, thermomagnetic iron oxides and docetaxel loaded nanoparticles of biodegradable polymers. Biomaterials 2012;33:7519-29.

- 67. Khaniabadi PM, Shahbazi-Gahrouei D, Aziz AA, Dheyab MA, Khaniabadi BM, Mehrdel B, et al. Trastuzumab conjugated porphyrin-superparamagnetic iron oxide nanoparticle: A potential PTT-MRI bimodal agent for herceptin positive breast cancer. Photodiagnosis Photodyn Ther 2020;31:101896.
- Maier-Hauff K, Ulrich F, Nestler D, Niehoff H, Wust P, Thiesen B, et al. Efficacy and safety of intratumoral thermotherapy using magnetic iron-oxide nanoparticles combined with external beam radiotherapy on patients with recurrent glioblastoma multiforme. J Neurooncol 2011;103:317-24.
- 69. Wu MR, Lee CH, Hsiao JK. Bidirectional enhancement of cell proliferation between iron oxide nanoparticle-labeled mesenchymal stem cells and choroid plexus in a cell-based therapy model of ischemic stroke. Int J Nanomedicine 2020;15:9181-95.
- Chien LY, Hsiao JK, Hsu SC, Yao M, Lu CW, Liu HM, et al. *In vivo* magnetic resonance imaging of cell tropism, trafficking mechanism, and therapeutic impact of human mesenchymal stem cells in a murine glioma model. Biomaterials 2011;32:3275-84.
- Chen CL, Siow TY, Chou CH, Lin CH, Lin MH, Chen YC, et al. Targeted superparamagnetic Iron OXIDE NANOPARTICLES for *in vivo* magnetic resonance imaging of T-cells in rheumatoid arthritis. Mol Imaging Biol 2017;19:233-44.
- 72. Tong W, Hui H, Shang W, Zhang Y, Tian F, Ma Q, et al. Highly sensitive magnetic particle imaging of vulnerable atherosclerotic plaque with active myeloperoxidase-targeted nanoparticles. Theranostics 2021;11:506-21.