Review Article



Role of adipocyte browning in prostate and breast tumor microenvironment

Hui-Chen Ku^a, Ching-Feng Cheng^{a, b, c}*

^aDepartment of Pediatrics, Taipei Tzu Chi Hospital, Buddhist Tzu Chi Medical Foundation, New Taipei, Taiwan, ^bInstitute of Biomedical Sciences, Academia Sinica, Taipei, Taiwan, ^cDepartment of Pediatrics, School of Medicine, Tzu Chi University, Hualien, Taiwan

ubmission	: 22-Mar-2022
evision	: 06-Apr-2022
cceptance	: 15-Apr-2022

Abstract

Prostate cancer (PC) and breast cancer (BC) are the most common cancers in men and women, respectively, in developed countries. The increased incidence of PC and BC largely reflects an increase in the prevalence of obesity and metabolic syndrome. In pathological conditions involving the development and progression of PC and BC, adipose tissue plays an important role via paracrine and endocrine signaling. The increase in the amount of local adipose tissue, specifically periprostatic adipose tissue, may be a key contributor to the PC pathobiology. Similarly, breast adipose tissue secretion affects various aspects of BC by influencing tumor progression, angiogenesis, metastasis, and microenvironment. In this context, the role of white adipose tissue (WAT) has been extensively studied. However, the influence of browning of the WAT on the development and progression of PC and BC is unclear and has received less attention. In this review, we highlight that adipose tissue plays a vital role in the regulation of the tumor microenvironment in PC or BC and highlight the probable underlying mechanisms linking adipose tissue with PC or BC. We further discuss whether the browning of WAT could be a therapeutic strategy for the treatment of PC and BC.

Keywords: Adipocyte browning, Breast cancer, Prostate cancer, Tumor microenvironment

Revision: 06-Apr-2022Acceptance: 15-Apr-2022Web Publication: 27-Jun-2022

INTRODUCTION

Si

 ${\cal P}^{
m rostate}$ cancer (PC) is the most common type of cancer in men in developed countries [1]. There is growing evidence to demonstrate the association between obesity and carcinoma aggressiveness, poor treatment outcomes, and a higher risk of cancer-specific mortality for PC [2-5]. Similarly, breast cancer (BC) is the most commonly diagnosed cancer and the main cause of cancer-related deaths in women worldwide. General and central obesity are risk factors for many chronic diseases [6,7] and are often defined by the body mass index (BMI) or waist-to-hip ratio (WHR) [8]. Overweight and obesity are associated with an elevated risk of 13 types of cancers [9,10]. The aim of this review was to explore the role of adipose tissue in the regulation of the tumor microenvironment in PC or BC by discussing the following: (1) the relationship between obesity and PC or BC, (2) possible physiological mechanisms linking obesity and the progression of PC or BC, and (3) white adipose tissue (WAT) browning as a potential therapeutic strategy for PC or BC via the improvement of tumor microenvironment.

OBESITY AND PROSTATE CANCER

Numerous studies have been conducted to understand how PC progression is affected by the consequences of an

Access this article online			
Quick Response Code:	Website: www.tcmjmed.com		
	DOI: 10.4103/tcmj.tcmj_62_22		

obese environment, such as increased systemic inflammation, hyperinsulinemia, altered adipokine profiles, and upregulated lipid availability [11,12]. Augmented synthesis and uptake of lipids are important hallmarks of PC and are modulated by androgen signaling (the key driver of PC pathogenesis) [13,14]. In addition, increased local adipose tissue amounts, specifically peri-prostatic adipose tissue (PPAT), may be associated with a higher grade or aggressiveness of PC; obesity-modulated alteration to the size of this lipid depot may be a key contributor to PC pathobiology. Obesity is a condition of chronic inflammation that is characterized by enhanced secretion of inflammatory cytokine, including interleukin (IL)-6, monocyte chemoattractant protein-1, and tumor necrosis factor- α (TNF- α), by adipose tissues [15] [Table 1]. These inflammatory cytokines are associated with PC progression both in clinical and in vitro studies [16-18]. Particularly, IL-6 secreted by PPAT in patients with PC demonstrated a concentration 375 times greater than that in the matched patient serum and was found to be associated significantly with the disease

*Address for correspondence: Dr. Ching-Feng Cheng, Department of Pediatrics, Taipei Tzu Chi Hospital, Buddhist Tzu Chi Medical Foundation, 289, Jianguo Road, Xindian District, New Taipei, Taiwan. E-mail: chengcf@mail.tcu.edu.tw

This is an open access journal, and articles are distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 4.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as appropriate credit is given and the new creations are licensed under the identical terms.

For reprints contact: WKHLRPMedknow_reprints@wolterskluwer.com

How to cite this article: Ku HC, Cheng CF. Role of adipocyte browning in prostate and breast tumor microenvironment. Tzu Chi Med J 2022;34(4):359-66.

pathological grade [19]. Moreover, PPAT inflammation, defined by the existence of crown-like structures (CLS), was found to be associated with larger adipocyte size, higher circulating levels of insulin and triglycerides, and highgrade PC [20]. Adipose tissues are endocrine organs that synthesize, secrete, and metabolize steroid hormones from circulating precursors. In addition, adipose tissues contain several androgens and androgen precursors, including testosterone, dihydrotestosterone, androstenedione, progesterone, and dehydroepiandrosterone [21], which, in the case of PPAT, supply a valid local extragonadal source of androgens that may reinforce PC growth and metastasis. PPAT also produces aromatase enzymes, which convert androgens to estrogens [22], and several studies suggest that estradiol is a modulator in PC pathogenesis and progression [23]. In addition, estrogen can activate both wild-type and mutated androgen receptors [24]. In summary, obesity is rather consistently associated with an increased risk of aggressive PC.

RECIPROCAL INTERACTIONS BETWEEN ADIPOSE TISSUE AND PROSTATE CANCER

The reciprocal interaction between adipocytes and tumor cells re-shapes adipocytes to a less differentiated condition referred to as cancer-associated adipocytes, a phenotype favorable to more aggressive tumors such as PC [22,25-27]. Several studies suggest that cancer-associated adipocytes can increase the malignant features of the cancer cells, eventually leading to detrimental positive feedback [25,28,29]. Culturing human PPAT using PC3 cell-derived conditioned medium (CM) increases the secretion of adipokines, TNF-a, IL-6, and osteopontin and enhances matrix metalloproteinase (MMP)-9 activity [25]. Furthermore, preadipocytes primed with PC CM undergo neoplastic-like transformation such as genetic instability, mesenchymal-to-epithelial transition, and formation of prostate-like neoplastic lesions in vivo [30]. PC is affected by adipocyte-secreted factors that increase the cells' ability to proliferate, migrate, and/or invade [19,26,29,31-34]. The biopsies of human prostate specimens or PPAT collected after prostatectomy showed a strong concentration gradient of the adipokine CCL7, suggesting that the PPAT secretome passively diffuses away from it into the tumor tissues to increase the directed migration of PC cells [26]. The CM of PPAT demonstrates higher MMP activity compared with that seen in peri-peritoneal visceral adipose tissue [29], which degrades the extracellular matrix proteins and promotes the invasion of cancer cells into the surrounding tissues [35]. Direct adipocyte-prostate cell crosstalk has been observed in their co-culture models. Mature rat epididymal adipocytes influenced the growth and differentiation of normal rat prostatic epithelium [36] or human PC [37,38] when cocultured in a three-dimensional collagen gel matrix. These effects were accompanied by an upregulated expression (20-fold) of the cytokines, including vascular endothelial growth factor and platelet-derived growth factor [37], and activation of the phosphatidylinositol 3-kinase (PI3K) pathway [38] in the PC3 cells. However, different studies have reported considerable variability - PPAT CM showed a stimulatory effect on PC3 and LNCaP cell migration in one

360

study [29], and coculturing rat epididymal adipocytes with PC3 cells increased PC3 proliferation in one study [37] – but these findings were contradicted in other studies [38]. This discrepancy is probably due to the differences in the nature of the cell lines and experimental methodologies used. The functional significance of adipocyte-PC cell interactions is emphasized by a study using a subcutaneous *in vivo* tumor model, in which larger tumors were generated by co-injection of PC cells with preadipocytes than by injection of only PC cells [39]. Thus, targeting the biological modulators of the tumor microenvironment, which links PPAT and PC, has the potential to reduce PC progression.

Possible mechanisms connecting obesity and prostate cancer

The mechanisms connecting adiposity and the progression of PC are poorly understood, and may be multifactorial [40]. In this respect, prospective components consisting of adipokine signaling pathways, sex hormone concentrations, and variation along the insulin/insulin-like-growth-factor (IGF) axis were involved [41,42]. (1) In adipokine signaling pathways, leptin and adiponectin are the two most plentiful and well-studied adipokines. High concentrations of adiponectin inhibit PC cell growth [43,44] [Table 2] and extend a beneficial effect on PC by suppressing inflammation,

Table 1: Adipokines with their alterations in obesity and					
beneficial/detrimental effects					
Adipokines	Alteration in	Beneficial or detrimental effects			
	obesity				
Adiponectin	Reduction	Anti-inflammation, insulin sensitizing			
Leptin	Increase due to	Modulates appetite and energy expenditure			
	leptin resistance				
Resistin	Increase	Induces insulin resistance, pro-inflammation			
TNF-α	Increase	Impairs the insulin signaling, contributes to			

		the pro-inflammatory state		
IL-6	Increase	Enhances C-reactive protein release from		
		the liver, causes insulin resistance, leads to		
		the pro-inflammatory state		
MCP-1	Increase	Leads to the pro-inflammatory state		
THE au Trum on a comparing factor of MCD 1. Man courts a home attractant				

TNF-α: Tumor necrosis factor-α, MCP-1: Monocyte chemoattractant protein-1, IL-6: Interleukin 6

Table 2: Effects of adipokines on prostate cancer and breast

cancer			
Adipokine	Tumor type	Cancer development	References
Leptin	PC	Increase	[43]
	BC	Increase	[45]
Adiponectin	PC	Decrease	[44]
	BC	Decrease	[46]
MCP-1	PC	Increase	[15]
	BC	Increase	[15]
TNF-α	PC	Increase	[25]
	BC	Increase	[47]
IL-6	PC	Increase	[19]
	BC	Increase	[46]

TNF-a: Tumor necrosis factor-a, MCP-1: Monocyte chemoattractant protein-1, IL-6: Interleukin 6, PC: Prostate cancer, BC: Breast cancer

activating fatty acid oxidation, ameliorating insulin sensitivity and glucose metabolism [44,48], and stimulating adenosine monophosphate-activated protein kinase (AMPK) activity [49]. Conversely, high concentrations of leptin have a pro-tumor effect in DU145 and PC-3 but not in LNCaP-FGC PC cell lines [32]. The relationship between these adipokines and PC progression needs to be elucidated [50]. (2) Sex hormone: Massillo et al. have found that estradiol induces the proliferation of androgen-sensitive cells, whereas it diminishes the proliferation of androgen-insensitive cell lines. Furthermore, high-fat diet (HFD)-fed mice had elevated concentrations of estradiol, which was associated with increased PC cell growth [51]. Although preclinical studies have demonstrated the link between estradiol and PC progression [52], further investigation is required to substantiate the results [53]. (3) Insulin and IGF-axis: Insulin resistance is strongly associated with obesity, leading to high insulin levels circulating in the blood [54]. Insulin increases cell proliferation and glucose consumption in PC cells but not in noncancerous prostate epithelial cells [55]. The IGF-axis is composed of cell surface receptors, ligands, IGF-binding proteins, and proteases [41]. Epidemiological studies have shown that higher serum IGF-1 concentrations and downregulated circulating IGFBP-3 levels are correlated with an increased risk of developing PC [56]. IGFBP-3 has been shown to induce apoptosis in a PC-3 cell line in vitro [57]. In addition, previous studies have indicated that exercise and nutrition interventions could decrease the BMI and weight loss may be advantageous in ameliorating IGFBP concentrations, leading to decreased bioavailable IGF-1 and reduced risk of PC progression [58,59]. In conclusion, the regulation of adipokine signaling, sex hormones, and insulin and IGF-axis in the tumor microenvironment may have the potential for reducing PC progression [Figure 1].

OBESITY AND BREAST CANCER

A meta-analysis of cohort studies demonstrated positive associations between BMI and WHR with obesity-related cancers, such as postmenopausal BC [60]. However, there is still disagreement on their influence on the risk of premenopausal BC, which may be caused by ethnic differences or/and sample size of the clinical study. Recently, a study on Korean women found that there was a negative association between obesity and BC in premenopausal women [61]. Nonetheless, several studies showed no remarkable effect of obesity on the risk of BC in Asian premenopausal women [62,63]. In addition, an increased risk of triple-negative BC was observed in obese type II (BMI \ge 30 kg/m²) premenopausal Korean women [64].

CROSSTALK BETWEEN ADIPOCYTES AND BREAST CANCER

Obesity is greatly related to a dysfunctional metabolism in adipocytes resulting in several chronic diseases. High levels of free fatty acids (FFA), cholesterol, glycerol, and triglycerides in serum impact breast tumor initiation, development, and migration [65-69]. In vitro coculture of mature adipocytes with BC cells enhances BC cell proliferation, which strongly suggests that adipocytes directly impact cancer cells by their secretions [70]. FFA are obtained from daily meals, which deposit as lipid droplets in the adipose tissue. Inflammation-induced obesity is an essential mechanism in the development and invasion of BC [66,67,71]. Saturated fatty acids reportedly activate toll-like receptor 4 to augment inflammation that leads to angiogenesis and tumor progression [72]. Inflamed microenvironment promotes adipocyte cell death, recruits macrophages, and leads to the formation of CLS [71]. The number of CLS is nine times higher in cancer patients with obesity than in lean women with BC and is often related to poor prognosis [73,74]. A study demonstrated that induced inflammation in WAT and increased CLS reduced the survival rate in patients [74]. Furthermore, saturated fatty acids can activate NF-kB, leading to TNF- α production, which affects BC cell proliferation, invasion, and metastasis [47]. FFAs induce BC invasion by activating the epidermal growth factor receptor, GTP-binding protein, and protein kinase C pathway [75], and controlling cell proliferation via PI3K [76] and cell migration through FFA receptor 1 and 4 and AKT pathway activation [77]. In addition, obesity-related factors within the tumor and the breast microenvironment are now known to regulate several important metabolic pathways: PI3K-RAC serine/threonine protein kinase (AKT), hypoxia-inducible factor 1α , liver kinase B1-AMPK, and p53. Dysregulated metabolic pathways in the breast microenvironment can support tumor growth. In summary, targeting the biological regulator of the tumor microenvironment between WAT and BC has the potential to decrease BC progression.

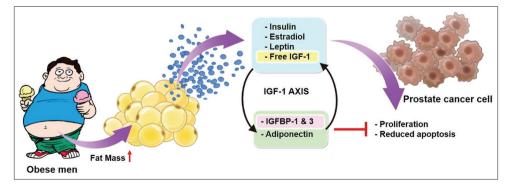


Figure 1: Proposed mechanisms for the link between obesity and prostate cancer progression. IGF: Insulin-like growth factor

Possible mechanisms linking obesity and breast cancer

The mechanism for the obesity-driven BC is very complex, and the underlying mechanisms in this process mainly consist of adipokines, insulin/IGF, sex hormone, and chronic inflammation; their dysregulation can enhance BC incidence and progression in the following ways. (1) Adipokines: Leptin-dependent secretion of the extracellular matrix proteins such as MMP-2 and MMP-9 and invasion in a FAK and Src-dependent manner suggest that leptin boosted the development of a more aggressive invasive phenotype in BC [45] [Table 2]. In addition, serum levels of adiponectin were decreased in a diet-induced obese mouse model, which was negatively correlated with obesity and increased the BC recurrence [46]. (2) Insulin and IGF: Insulin in conjunction with inflammation can enhance BC growth and metastasis [78]. Furthermore, HFD-fed obese mice showed hyperinsulinemia, upregulated IGF-1 levels, and accelerated BC recurrence, suggesting that the insulin/ IGF-1 signaling pathway is a potential regulator for obesity and BC recurrence [46]. (3) Sex hormone: Aromatase, a rate-limiting enzyme, is secreted by stromal cells of adipose tissue that converts androstenedione to estrone, subsequently forming estrogen after menopause. Aromatase upregulation in the breast tissue of obese patients resulted in an increased risk of hormone receptor-positive BC in obese postmenopausal women [79]. (4) Chronic inflammation: Studies have found that obesity decreases the local IL-10 levels in the mammary fat pad of ovariectomized mice, resulting in the upregulation of aromatase and leading to BC progression [80]. In summary, targeting of adipokines, insulin/IGF-1, sex hormones, and inflammation in the BC tumor microenvironment may have the potential to hinder BC progression [Figure 2].

Factors involved in white-to-brown adipocyte conversion

There are two different types of adipose tissues – WAT, cells of which contain a large single, spherical lipid vacuoles and few mitochondria, and brown adipose tissue (BAT), cells of which contain small and multilocular lipid droplets and large number of mitochondria. BAT is the principal effector organ of nonshivering thermogenesis and can use a large amount of glucose and lipid from circulation to promote negative energy balance. Hence, it

will induce thermogenesis, dissipate heat, improve glucose metabolism, and develop insulin resistance in obese individuals [81-84]. Therefore, BAT is now known to exert anti-type 2 diabetes effects associated with improvement of dyslipidemia and decreased insulin resistance [85-88]. The metabolic adaptations during white-to-brown adipocyte conversion are not well known. Several studies have shown that another type of brown cells, known as the beige or brite (brown in white) cells, exists in both mouse and human [89-91]. Beige cells are generated postnatally within WAT in response to cold or adrenergic stimulations. Both classical brown fat and beige cells are rich in mitochondria and uniquely express UCP1. Although both share the same thermogenic function, they arise from entirely different cell lineages [89,92]. In contrast to brown cells that express both myogenic genes Myf5 and Pax7 [93,94], beige cells are generated postnatally in WAT depots and arise from Myf5-precursor lineage that expresses PDGFRa [89,95,96] or through transdifferentiation of mature white [96-98] adipocytes in response to cold or β -adrenergic stimulation. In human, beige adipocytes have been observed in white fat depots [87,99]. Morphological and histological data indicate the presence of cells with an intermediate phenotype, suggesting that conversion of white into beige adipocytes likely occurs [87]. There are three main transcriptional regulators of classical BAT development, namely PR domain containing 16 (PRDM16), peroxisome proliferator-activated receptor y, and peroxisome proliferator-activated receptor γ coactivator 1 α , which are key nodes in the regulation of inducible brown fat. In addition, some transcription factors and coregulators are involved in the browning process of WAT such as forkhead box protein C2 [100], steroid receptor coactivator-1, transcriptional intermediary factor-2 [101], T-box 15 (TBX15) [102], and mitochondrial transcription factor A [103]. Moreover, secreted proteins, including irisin [104], FGF21 [105], cardiac natriuretic peptide [106], and bone morphogenetic protein 7 [107], had been reported regulate white-to-brown conversion. Furthermore, to different components of the immune system have been reported to promote browning, such as eosinophils [108], macrophages [109.110] and ILC2s [111.112]; several cytokines are involved in the regulation of browning. In sum, targeting transcription factors and coregulators involved in the browning of WAT may have the potential to combat obesity or improve the tumor microenvironment and needs further investigation.

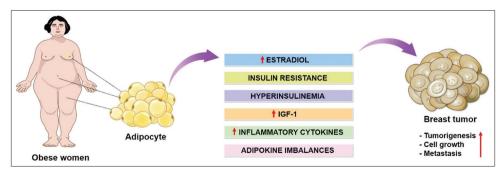


Figure 2: Proposed mechanisms for the association between obesity and breast cancer progression. IGF: Insulin-like growth factor

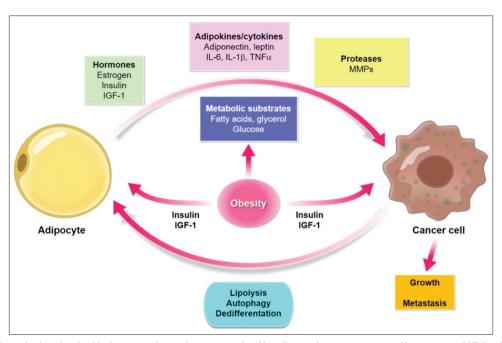


Figure 3: The possible mechanisms involved in the tumor microenvironment regulated by adipocyte in prostate cancer and breast cancer. IGF: Insulin-like growth factor, IL: Interleukin, TNF: Tumor necrosis factor, MMP: Matrix metalloproteinase

THERAPEUTIC BROWNING OF WHITE ADIPOSE TISSUE IN THE TUMOR MICROENVIRONMENT

In pathological conditions such as the development and progression of PC and BC, adipose tissue plays an important role via paracrine and endocrine signaling. Although implied, the influence of WAT browning on the development and progression of PC and BC is unclear and has received less attention. Browning of WAT can be achieved with benzyl isothiocyanate and Honokiol, which increase the expression of BAT marker genes (UCP1, PRDM16, EVOL3, COX7a, and CIDEA) in WAT. It not only abolishes the pro-cancer effects of WAT on BC cells but also changes the secretome profile of WAT [113]. Furthermore, the in vitro and in vivo models with primary brown adipose cells (BACs) indicate that primary BACs can directly decrease the viability of H22 cells, a hepatocellular carcinoma cell line, and the growth of tumors. In conclusion, BACs may be a potential therapeutic tool for the treatment of hepatocellular carcinoma [114]. However, whether browning WAT could be a therapeutic strategy for the treatment of PC and BC needs further examination.

CONCLUSION

Epidemiological and clinical evidence has shown a consistent association between obesity with cancer progression and increased mortality of PC and BC patients. However, the underlying mechanisms linking them remain unclear. Clinical studies found that alteration of insulin and IGF-axis, sex hormone concentrations, and adipokine signaling can increase cancer cell proliferation in individuals with obesity. In addition, these factors can also synthetically affect angiogenesis, oncogene activation, immune cell dysfunction, and oxidative stress, which can modulate the behaviors of PC or BC cells and tumor microenvironment. Targeting adipocyte-derived molecules may be a potential therapeutic approach to ameliorate the prognosis of obese patients. Furthermore, a thorough understanding of the physiological mechanisms of obesity on treatment effectiveness and tolerance is necessary for improving the efficacy of PC or BC therapy. A summary of the possible mechanisms involved in the tumor microenvironment regulated by adipocyte in PC and BC is depicted in Figure 3. Thus, it can be concluded that regulation of adipocyte function is a novel therapeutic strategy for the treatment of PC or BC.

Financial support and sponsorship

This work was supported by the Ministry of Science and Technology (MOST 110-2314-B-303-011-MY3 to C-FC), Buddhist Tzu Chi Medical Foundation (TCMMP111-01-02 to C-FC), and Tzu Chi and Academia Sinica cooperation (TCAS-108-01 translational research grants to C-FC).

Conflicts of interest

Dr. Ching-Feng Cheng, an editorial board member at *Tzu Chi Medical Journal*, had no role in the peer review process of or decision to publish this article. The other author declared no conflicts of interest in writing this article.

References

- Zhou CK, Check DP, Lortet-Tieulent J, Laversanne M, Jemal A, Ferlay J, et al. Prostate cancer incidence in 43 populations worldwide: An analysis of time trends overall and by age group. Int J Cancer 2016;138:1388-400.
- Zhong S, Yan X, Wu Y, Zhang X, Chen L, Tang J, et al. Body mass index and mortality in prostate cancer patients: A dose-response meta-analysis. Prostate Cancer Prostatic Dis 2016;19:122-31.
- Hu MB, Liu SH, Jiang HW, Bai PD, Ding Q. Obesity affects the biopsy-mediated detection of prostate cancer, particularly high-grade prostate cancer: A dose-response meta-analysis of 29,464 patients. PLoS One 2014;9:e106677.
- 4. Arnold M, Karim-Kos HE, Coebergh JW, Byrnes G, Antilla A, Ferlay J, et al. Recent trends in incidence of five common cancers in 26 European

countries since 1988: Analysis of the European Cancer Observatory. Eur J Cancer 2015;51:1164-87.

- Arnold M, Leitzmann M, Freisling H, Bray F, Romieu I, Renehan A, et al. Obesity and cancer: An update of the global impact. Cancer Epidemiol 2016;41:8-15.
- Upadhyay J, Farr O, Perakakis N, Ghaly W, Mantzoros C. Obesity as a disease. Med Clin North Am 2018;102:13-33.
- Avgerinos KI, Spyrou N, Mantzoros CS, Dalamaga M. Obesity and cancer risk: Emerging biological mechanisms and perspectives. Metabolism 2019;92:121-35.
- Kanazawa M, Yoshiike N, Osaka T, Numba Y, Zimmet P, Inoue S. Criteria and classification of obesity in Japan and Asia-Oceania. World Rev Nutr Diet 2005;94:1-12.
- Arnold M, Pandeya N, Byrnes G, Renehan PA, Stevens GA, Ezzati PM, et al. Global burden of cancer attributable to high body-mass index in 2012: A population-based study. Lancet Oncol 2015;16:36-46.
- Lauby-Secretan B, Scoccianti C, Loomis D, Grosse Y, Bianchini F, Straif K, et al. Body fatness and cancer-viewpoint of the IARC Working Group. N Engl J Med 2016;375:794-8.
- 11. Zadra G, Photopoulos C, Loda M. The fat side of prostate cancer. Biochim Biophys Acta 2013;1831:1518-32.
- Grossmann M, Wittert G. Androgens, diabetes and prostate cancer. Endocr Relat Cancer 2012;19:F47-62.
- Butler LM, Centenera MM, Swinnen JV. Androgen control of lipid metabolism in prostate cancer: Novel insights and future applications. Endocr Relat Cancer 2016;23:R219-27.
- 14. Hanahan D. Hallmarks of cancer: New dimensions. Cancer Discov 2022;12:31-46.
- Divella R, De Luca R, Abbate I, Naglieri E, Daniele A. Obesity and cancer: The role of adipose tissue and adipo-cytokines-induced chronic inflammation. J Cancer 2016;7:2346-59.
- Yu D, Zhong Y, Li X, Li Y, Li X, Cao J, et al. ILs-3, 6 and 11 increase, but ILs-10 and 24 decrease stemness of human prostate cancer cells *in vitro*. Oncotarget 2015;6:42687-703.
- Sharma J, Gray KP, Harshman LC, Evan C, Nakabayashi M, Fichorova R, et al. Elevated IL-8, TNF-α, and MCP-1 in men with metastatic prostate cancer starting androgen-deprivation therapy (ADT) are associated with shorter time to castration-resistance and overall survival. Prostate 2014;74:820-8.
- Xu H, Hu MB, Bai PD, Zhu WH, Liu SH, Hou JY, et al. Proinflammatory cytokines in prostate cancer development and progression promoted by high-fat diet. Biomed Res Int 2015;2015:249741.
- Finley DS, Calvert VS, Inokuchi J, Lau A, Narula N, Petricoin EF, et al. Periprostatic adipose tissue as a modulator of prostate cancer aggressiveness. J Urol 2009;182:1621-7.
- Gucalp A, Iyengar NM, Zhou XK, Giri DD, Falcone DJ, Wang H, et al. Periprostatic adipose inflammation is associated with high-grade prostate cancer. Prostate Cancer Prostatic Dis 2017;20:418-23.
- Blouin K, Veilleux A, Luu-The V, Tchernof A. Androgen metabolism in adipose tissue: Recent advances. Mol Cell Endocrinol 2009;301:97-103.
- Nieman KM, Romero IL, Van Houten B, Lengyel E. Adipose tissue and adipocytes support tumorigenesis and metastasis. Biochim Biophys Acta 2013;1831:1533-41.
- Di Zazzo E, Galasso G, Giovannelli P, Di Donato M, Di Santi A, Cernera G, et al. Prostate cancer stem cells: The role of androgen and estrogen receptors. Oncotarget 2016;7:193-208.
- Susa T, Ikaga R, Kajitani T, Iizuka M, Okinaga H, Tamamori-Adachi M, et al. Wild-type and specific mutant androgen receptor mediates transcription via 17β-estradiol in sex hormone-sensitive cancer cells. J Cell Physiol 2015;230:1594-606.
- 25. Ribeiro RJ, Monteiro CP, Cunha VF, Azevedo AS, Oliveira MJ, Monteiro R, et al. Tumor cell-educated periprostatic adipose tissue acquires an aggressive cancer-promoting secretory profile. Cell Physiol

Biochem 2012;29:233-40.

- Laurent V, Guérard A, Mazerolles C, Le Gonidec S, Toulet A, Nieto L, et al. Periprostatic adipocytes act as a driving force for prostate cancer progression in obesity. Nat Commun 2016;7:10230.
- Dirat B, Bochet L, Dabek M, Daviaud D, Dauvillier S, Majed B, et al. Cancer-associated adipocytes exhibit an activated phenotype and contribute to breast cancer invasion. Cancer Res 2011;71:2455-65.
- Ribeiro R, Monteiro C, Catalán V, Hu P, Cunha V, Rodríguez A, et al. Obesity and prostate cancer: Gene expression signature of human periprostatic adipose tissue. BMC Med 2012;10:108.
- Ribeiro R, Monteiro C, Cunha V, Oliveira MJ, Freitas M, Fraga A, et al. Human periprostatic adipose tissue promotes prostate cancer aggressiveness *in vitro*. J Exp Clin Cancer Res 2012;31:32.
- Abd Elmageed ZY, Yang Y, Thomas R, Ranjan M, Mondal D, Moroz K, et al. Neoplastic reprogramming of patient-derived adipose stem cells by prostate cancer cell-associated exosomes. Stem Cells 2014;32:983-97.
- Moreira Â, Pereira SS, Costa M, Morais T, Pinto A, Fernandes R, et al. Adipocyte secreted factors enhance aggressiveness of prostate carcinoma cells. PLoS One 2015;10:e0123217.
- Onuma M, Bub JD, Rummel TL, Iwamoto Y. Prostate cancer cell-adipocyte interaction: Leptin mediates androgen-independent prostate cancer cell proliferation through c-Jun NH2-terminal kinase. J Biol Chem 2003;278:42660-7.
- Sacca PA, Creydt VP, Choi H, Mazza ON, Fletcher SJ, Vallone VB, et al. Human periprostatic adipose tissue: Its influence on prostate cancer cells. Cell Physiol Biochem 2012;30:113-22.
- 34. Ito Y, Ishiguro H, Kobayashi N, Hasumi H, Watanabe M, Yao M, et al. Adipocyte-derived monocyte chemotactic protein-1 (MCP-1) promotes prostate cancer progression through the induction of MMP-2 activity. Prostate 2015;75:1009-19.
- Duffy MJ, Maguire TM, Hill A, McDermott E, O'Higgins N. Metalloproteinases: Role in breast carcinogenesis, invasion and metastasis. Breast Cancer Res 2000;2:252-7.
- Tokuda Y, Toda S, Masaki Z, Sugihara H. Proliferation and differentiation of rat dorsal prostatic epithelial cells in collagen gel matrix culture, focusing upon effects of adipocytes. Int J Urol 1999;6:509-19.
- Tokuda Y, Satoh Y, Fujiyama C, Toda S, Sugihara H, Masaki Z. Prostate cancer cell growth is modulated by adipocyte-cancer cell interaction. BJU Int 2003;91:716-20.
- Kaneko A, Satoh Y, Tokuda Y, Fujiyama C, Udo K, Uozumi J. Effects of adipocytes on the proliferation and differentiation of prostate cancer cells in a 3-D culture model. Int J Urol 2010;17:369-76.
- Prantl L, Muehlberg F, Navone NM, Song YH, Vykoukal J, Logothetis CJ, et al. Adipose tissue-derived stem cells promote prostate tumor growth. Prostate 2010;70:1709-15.
- 40. Deng T, Lyon CJ, Bergin S, Caligiuri MA, Hsueh WA. Obesity, inflammation, and cancer. Annu Rev Pathol 2016;11:421-49.
- 41. Adesunloye BA. Mechanistic insights into the link between obesity and prostate cancer. Int J Mol Sci 2021;22:3935.
- 42. Allott EH, Masko EM, Freedland SJ. Obesity and prostate cancer: Weighing the evidence. Eur Urol 2013;63:800-9.
- Bub JD, Miyazaki T, Iwamoto Y. Adiponectin as a growth inhibitor in prostate cancer cells. Biochem Biophys Res Commun 2006;340:1158-66.
- Tahergorabi Z, Khazaei M, Moodi M, Chamani E. From obesity to cancer: A review on proposed mechanisms. Cell Biochem Funct 2016;34:533-45.
- 45. Juárez-Cruz JC, Zuñiga-Eulogio MD, Olea-Flores M, Castañeda-Saucedo E, Mendoza-Catalán MÁ, Ortuño-Pineda C, et al. Leptin induces cell migration and invasion in a FAK-Src-dependent manner in breast cancer cells. Endocr Connect 2019;8:1539-52.
- Ecker BL, Lee JY, Sterner CJ, Solomon AC, Pant DK, Shen F, et al. Impact of obesity on breast cancer recurrence and minimal residual disease. Breast Cancer Res 2019;21:41.
- 47. Subbaramaiah K, Howe LR, Bhardwaj P, Du B, Gravaghi C, Yantiss RK,

et al. Obesity is associated with inflammation and elevated aromatase expression in the mouse mammary gland. Cancer Prev Res (Phila) 2011;4:329-46.

- Li H, Stampfer MJ, Mucci L, Rifai N, Qiu W, Kurth T, et al. A 25-year prospective study of plasma adiponectin and leptin concentrations and prostate cancer risk and survival. Clin Chem 2010;56:34-43.
- Champ CE, Francis L, Klement RJ, Dickerman R, Smith RP. Fortifying the treatment of prostate cancer with physical activity. Prostate Cancer 2016;2016:9462975.
- Burton AJ, Gilbert R, Tilling K, Langdon R, Donovan JL, Holly JM, et al. Circulating adiponectin and leptin and risk of overall and aggressive prostate cancer: A systematic review and meta-analysis. Sci Rep 2021;11:320.
- Massillo C, Dalton GN, Porretti J, Scalise GD, Farré PL, Piccioni F, et al. CTBP1/CYP19A1/estradiol axis together with adipose tissue impacts over prostate cancer growth associated to metabolic syndrome. Int J Cancer 2019;144:1115-27.
- Bonkhoff H, Berges R. The evolving role of oestrogens and their receptors in the development and progression of prostate cancer. Eur Urol 2009;55:533-42.
- Yao S, Till C, Kristal AR, Goodman PJ, Hsing AW, Tangen CM, et al. Serum estrogen levels and prostate cancer risk in the prostate cancer prevention trial: A nested case-control study. Cancer Causes Control 2011;22:1121-31.
- Ahmadi H, Daneshmand S. Androgen deprivation therapy: Evidence-based management of side effects. BJU Int 2013;111:543-8.
- 55. Heidegger I, Ofer P, Doppler W, Rotter V, Klocker H, Massoner P. Diverse functions of IGF/insulin signaling in malignant and noncancerous prostate cells: Proliferation in cancer cells and differentiation in noncancerous cells. Endocrinology 2012;153:4633-43.
- Chan JM, Stampfer MJ, Giovannucci E, Gann PH, Ma J, Wilkinson P, et al. Plasma insulin-like growth factor-I and prostate cancer risk: A prospective study. Science 1998;279:563-6.
- 57. Rajah R, Valentinis B, Cohen P. Insulin-like growth factor (IGF)-binding protein-3 induces apoptosis and mediates the effects of transforming growth factor-beta1 on programmed cell death through a p53- and IGF-independent mechanism. J Biol Chem 1997;272:12181-8.
- Barnard RJ, Ngo TH, Leung PS, Aronson WJ, Golding LA. A low-fat diet and/or strenuous exercise alters the IGF axis *in vivo* and reduces prostate tumor cell growth *in vitro*. Prostate 2003;56:201-6.
- Wright JL, Plymate S, D'Oria-Cameron A, Bain C, Haugk K, Xiao L, et al. A study of caloric restriction versus standard diet in overweight men with newly diagnosed prostate cancer: A randomized controlled trial. Prostate 2013;73:1345-51.
- 60. Freisling H, Arnold M, Soerjomataram I, O'Doherty MG, Ordóñez-Mena JM, Bamia C, et al. Comparison of general obesity and measures of body fat distribution in older adults in relation to cancer risk: Meta-analysis of individual participant data of seven prospective cohorts in Europe. Br J Cancer 2017;116:1486-97.
- Park JW, Han K, Shin DW, Yeo Y, Chang JW, Yoo JE, et al. Obesity and breast cancer risk for pre- and postmenopausal women among over 6 million Korean women. Breast Cancer Res Treat 2021;185:495-506.
- 62. Cao S, Zhou J, Zhu Z, Wei F, Li W, Lu S, et al. Adult weight change and the risk of pre- and postmenopausal breast cancer in the Chinese Wuxi Exposure and Breast Cancer Study. Breast Cancer Res Treat 2019;173:647-55.
- Parr CL, Batty GD, Lam TH, Barzi F, Fang X, Ho SC, et al. Body-mass index and cancer mortality in the Asia-Pacific Cohort Studies Collaboration: Pooled analyses of 424,519 participants. Lancet Oncol 2010;11:741-52.
- 64. Jeong SH, An Y, Ahn C, Park B, Lee MH, Noh DY, et al. Body mass index and risk of breast cancer molecular subtypes in Korean women: A case-control study. Breast Cancer Res Treat 2020;179:459-70.

- Baek AE, Nelson ER. The contribution of cholesterol and its metabolites to the pathophysiology of breast cancer. Horm Cancer 2016;7:219-28.
- 66. Garcia-Estevez L, Moreno-Bueno G. Updating the role of obesity and cholesterol in breast cancer. Breast Cancer Res 2019;21:35.
- Gilbert CA, Slingerland JM. Cytokines, obesity, and cancer: New insights on mechanisms linking obesity to cancer risk and progression. Annu Rev Med 2013;64:45-57.
- Rose DP, Connolly JM. Effects of fatty acids and inhibitors of eicosanoid synthesis on the growth of a human breast cancer cell line in culture. Cancer Res 1990;50:7139-44.
- Monk JM, Turk HF, Liddle DM, De Boer AA, Power KA, Ma DW, et al. n-3 polyunsaturated fatty acids and mechanisms to mitigate inflammatory paracrine signaling in obesity-associated breast cancer. Nutrients 2014;6:4760-93.
- Rajarajan D, Selvarajan S, Charan Raja MR, Kar Mahapatra S, Kasiappan R. Genome-wide analysis reveals miR-3184-5p and miR-181c-3p as a critical regulator for adipocytes-associated breast cancer. J Cell Physiol 2019;234:17959-74.
- Ackerman SE, Blackburn OA, Marchildon F, Cohen P. Insights into the link between obesity and cancer. Curr Obes Rep 2017;6:195-203.
- Vigushin DM, Dong Y, Inman L, Peyvandi N, Alao JP, Sun C, et al. The nuclear oxysterol receptor LXRalpha is expressed in the normal human breast and in breast cancer. Med Oncol 2004;21:123-31.
- Morris PG, Hudis CA, Giri D, Morrow M, Falcone DJ, Zhou XK, et al. Inflammation and increased aromatase expression occur in the breast tissue of obese women with breast cancer. Cancer Prev Res (Phila) 2011;4:1021-9.
- Iyengar NM, Ghossein RA, Morris LG, Zhou XK, Kochhar A, Morris PG, et al. White adipose tissue inflammation and cancer-specific survival in patients with squamous cell carcinoma of the oral tongue. Cancer 2016;122:3794-802.
- Soto-Guzman A, Navarro-Tito N, Castro-Sanchez L, Martinez-Orozco R, Salazar EP. Oleic acid promotes MMP-9 secretion and invasion in breast cancer cells. Clin Exp Metastasis 2010;27:505-15.
- Hardy S, Langelier Y, Prentki M. Oleate activates phosphatidylinositol 3-kinase and promotes proliferation and reduces apoptosis of MDA-MB-231 breast cancer cells, whereas palmitate has opposite effects. Cancer Res 2000;60:6353-8.
- Marcial-Medina C, Ordoñez-Moreno A, Gonzalez-Reyes C, Cortes-Reynosa P, Perez Salazar E. Oleic acid induces migration through a FFAR1/4, EGFR and AKT-dependent pathway in breast cancer cells. Endocr Connect 2019;8:252-65.
- Valentino E, Bellazzo A, Di Minin G, Sicari D, Apollonio M, Scognamiglio G, et al. Mutant p53 potentiates the oncogenic effects of insulin by inhibiting the tumor suppressor DAB2IP. Proc Natl Acad Sci U S A 2017;114:7623-8.
- 79. Gucalp A, Zhou XK, Cook ED, Garber JE, Crew KD, Nangia JR, et al. A randomized multicenter phase II study of docosahexaenoic acid in patients with a history of breast cancer, premalignant lesions, or benign breast disease. Cancer Prev Res (Phila) 2018;11:203-14.
- Martínez-Chacón G, Brown KA, Docanto MM, Kumar H, Salminen S, Saarinen N, et al. IL-10 suppresses TNF-α-induced expression of human aromatase gene in mammary adipose tissue. FASEB J 2018;32:3361-70.
- Guerra C, Koza RA, Yamashita H, Walsh K, Kozak LP. Emergence of brown adipocytes in white fat in mice is under genetic control. Effects on body weight and adiposity. J Clin Invest 1998;102:412-20.
- Almind K, Manieri M, Sivitz WI, Cinti S, Kahn CR. Ectopic brown adipose tissue in muscle provides a mechanism for differences in risk of metabolic syndrome in mice. Proc Natl Acad Sci U S A 2007;104:2366-71.
- 83. Veyrat-Durebex C, Montet X, Vinciguerra M, Gjinovci A, Meda P, Foti M, et al. The Lou/C rat: A model of spontaneous food restriction associated with improved insulin sensitivity and decreased lipid storage in

adipose tissue. Am J Physiol Endocrinol Metab 2009;296:E1120-32.

- 84. Harms M, Seale P. Brown and beige fat: Development, function and therapeutic potential. Nat Med 2013;19:1252-63.
- de Souza CJ, Hirshman MF, Horton ES. CL-316,243, a beta3-specific adrenoceptor agonist, enhances insulin-stimulated glucose disposal in nonobese rats. Diabetes 1997;46:1257-63.
- Liu X, Pérusse F, Bukowiecki LJ. Mechanisms of the antidiabetic effects of the beta 3-adrenergic agonist CL-316243 in obese Zucker-ZDF rats. Am J Physiol 1998;274:R1212-9.
- Frontini A, Vitali A, Perugini J, Murano I, Romiti C, Ricquier D, et al. White-to-brown transdifferentiation of omental adipocytes in patients affected by pheochromocytoma. Biochim Biophys Acta 2013;1831:950-9.
- Peirce V, Vidal-Puig A. Regulation of glucose homoeostasis by brown adipose tissue. Lancet Diabetes Endocrinol 2013;1:353-60.
- Wu J, Boström P, Sparks LM, Ye L, Choi JH, Giang AH, et al. Beige adipocytes are a distinct type of thermogenic fat cell in mouse and human. Cell 2012;150:366-76.
- Ishibashi J, Seale P. Medicine. Beige can be slimming. Science 2010;328:1113-4.
- 91. Petrovic N, Walden TB, Shabalina IG, Timmons JA, Cannon B, Nedergaard J. Chronic peroxisome proliferator-activated receptor gamma (PPARgamma) activation of epididymally derived white adipocyte cultures reveals a population of thermogenically competent, UCP1-containing adipocytes molecularly distinct from classic brown adipocytes. J Biol Chem 2010;285:7153-64.
- 92. Rosen ED, Spiegelman BM. What we talk about when we talk about fat. Cell 2014;156:20-44.
- Seale P, Bjork B, Yang W, Kajimura S, Chin S, Kuang S, et al. PRDM16 controls a brown fat/skeletal muscle switch. Nature 2008;454:961-7.
- Lepper C, Fan CM. Inducible lineage tracing of Pax7-descendant cells reveals embryonic origin of adult satellite cells. Genesis 2010;48:424-36.
- Schulz TJ, Huang TL, Tran TT, Zhang H, Townsend KL, Shadrach JL, et al. Identification of inducible brown adipocyte progenitors residing in skeletal muscle and white fat. Proc Natl Acad Sci U S A 2011;108:143-8.
- Wang QA, Tao C, Gupta RK, Scherer PE. Tracking adipogenesis during white adipose tissue development, expansion and regeneration. Nat Med 2013;19:1338-44.
- Lee YH, Petkova AP, Konkar AA, Granneman JG. Cellular origins of cold-induced brown adipocytes in adult mice. FASEB J 2015;29:286-99.
- Rosenwald M, Perdikari A, Rülicke T, Wolfrum C. Bi-directional interconversion of brite and white adipocytes. Nat Cell Biol 2013;15:659-67.
- Sidossis LS, Porter C, Saraf MK, Børsheim E, Radhakrishnan RS, Chao T, et al. Browning of subcutaneous white adipose tissue in humans after severe adrenergic stress. Cell Metab 2015;22:219-27.
- Cederberg A, Grønning LM, Ahrén B, Taskén K, Carlsson P, Enerbäck S. FOXC2 is a winged helix gene that counteracts obesity,

hypertriglyceridemia, and diet-induced insulin resistance. Cell 2001;106:563-73.

- Picard F, Géhin M, Annicotte J, Rocchi S, Champy MF, O'Malley BW, et al. SRC-1 and TIF2 control energy balance between white and brown adipose tissues. Cell 2002;111:931-41.
- 102. Gburcik V, Cawthorn WP, Nedergaard J, Timmons JA, Cannon B. An essential role for Tb×15 in the differentiation of brown and "brite" but not white adipocytes. Am J Physiol Endocrinol Metab 2012;303:E1053-60.
- 103. Vernochet C, Mourier A, Bezy O, Macotela Y, Boucher J, Rardin MJ, et al. Adipose-specific deletion of TFAM increases mitochondrial oxidation and protects mice against obesity and insulin resistance. Cell Metab 2012;16:765-76.
- 104. Boström P, Wu J, Jedrychowski MP, Korde A, Ye L, Lo JC, et al. A PGC1-α-dependent myokine that drives brown-fat-like development of white fat and thermogenesis. Nature 2012;481:463-8.
- 105. Fisher FM, Kleiner S, Douris N, Fox EC, Mepani RJ, Verdeguer F, et al. FGF21 regulates PGC-1α and browning of white adipose tissues in adaptive thermogenesis. Genes Dev 2012;26:271-81.
- 106. Bordicchia M, Liu D, Amri EZ, Ailhaud G, Dessi-Fulgheri P, Zhang C, et al. Cardiac natriuretic peptides act via p38 MAPK to induce the brown fat thermogenic program in mouse and human adipocytes. J Clin Invest 2012;122:1022-36.
- 107. Tseng YH, Kokkotou E, Schulz TJ, Huang TL, Winnay JN, Taniguchi CM, et al. New role of bone morphogenetic protein 7 in brown adipogenesis and energy expenditure. Nature 2008;454:1000-4.
- 108. Qiu Y, Nguyen KD, Odegaard JI, Cui X, Tian X, Locksley RM, et al. Eosinophils and type 2 cytokine signaling in macrophages orchestrate development of functional beige fat. Cell 2014;157:1292-308.
- 109. Fabbiano S, Suárez-Zamorano N, Rigo D, Veyrat-Durebex C, Stevanovic Dokic A, Colin DJ, et al. Caloric restriction leads to browning of white adipose tissue through type 2 immune signaling. Cell Metab 2016;24:434-46.
- 110. Nguyen KD, Qiu Y, Cui X, Goh YP, Mwangi J, David T, et al. Alternatively activated macrophages produce catecholamines to sustain adaptive thermogenesis. Nature 2011;480:104-8.
- 111. Brestoff JR, Kim BS, Saenz SA, Stine RR, Monticelli LA, Sonnenberg GF, et al. Group 2 innate lymphoid cells promote beiging of white adipose tissue and limit obesity. Nature 2015;519:242-6.
- 112. Lee MW, Odegaard JI, Mukundan L, Qiu Y, Molofsky AB, Nussbaum JC, et al. Activated type 2 innate lymphoid cells regulate beige fat biogenesis. Cell 2015;160:74-87.
- 113. Muniraj N, Siddharth S, Parida S, Shriver M, Nagalingam A, Sharma D. Abstract 2690: Therapeutic browning of white adipose tissue in the tumor microenvironment to inhibit breast cancer progression. Cancer Res 2021;81:2690.
- 114. Liu D, Li Y, Shang Y, Wang W, Chen SZ. Effect of brown adipose tissue/ cells on the growth of mouse hepatocellular carcinoma *in vitro* and *in vivo*. Oncol Lett 2019;17:3203-10.