Review Article



Indocyanine green: An old drug with novel applications

Chia-Hung Lua, Jong-Kai Hsiaoa,b*

^aDepartment of Medical Imaging, Taipei Tzu Chi Hospital, Buddhist Tzu Chi Medical Foundation, New Taipei, Taiwan, ^bSchool of Medicine, Tzu Chi University, Hualien, Taiwan

ABSTRACT

Indocyanine green (ICG), a US Food and Drug Administration-approved fluorescent compound, has been on the medical stage for more than 60 years. Current uses include hepatic function evaluation before surgical procedure and fundus evaluation. The large safety margin and near-infrared fluorescent optical advantage of the drug have proved useful in several clinical trials of intraoperative systems for tumor removal. Several nanoparticle-sized formulations for thermal ablation and photodynamic therapy have also been evaluated in animal experiments. Studies have attempted to manipulate ICG as a reporter fluorophore with initial success. In this article, we reviewed ICG's histological applications, chemical and physical properties, current clinical applications, ongoing clinical trials, and biomedical studies and prospects. We believe that ICG could be used with novel biotechnological techniques, such as fluorescent endoscopy and photoacoustic equipment, in a range of biomedical fields.

KEYWORDS: Fluorescent dye, Indocyanine green, Near-infrared, Optical Imaging

 Submission
 : 14-Aug-2020

 Revision
 : 05-Oct-2020

 Acceptance
 : 31-Oct-2020

 Web Publication
 : 01-Apr-2021

Introduction

11) the rising cost of new drug development and the growing concerns of the new drug's adverse reactions, interest has been focusing on the medicines that have been in the pharmaceutical markets for decades. Aspirin is one of the most well-known examples. People use aspirin since antiquity for relieving pain and fever. However, its application in cancer treatment and cardiovascular prevention is still ongoing [1-3]. Indocyanine green (ICG) is another less-known drug that stands in the medical field for 60 years. The optical fluorescence characteristic of ICG made its first launch in the medical field. As the imaging technique progresses into emerges. ICG along with novel imaging technique has been applied in diagnostic medical imaging, surgical procedure guidance, and new cancer therapy strategy. This review discusses the physical, chemical, and biological characteristics of ICG and its current usage in the clinical field, ongoing trials, and the potential applications proven in animal studies.

HISTORY OF INDOCYANINE GREEN IN BIOMEDICINE

ICG was developed by Kodak during World War II for color imaging purposes. The medical applications of ICG were approved by the United States Food and Drug Administration in 1959. In 1960, Fox reported the characteristics of ICG and the results of use in the Mayo Clinics [4]. ICG was originally, and is still, used to determine liver function. In 1963, Walker

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Quick Response Code:	
	Website: www.tcmjmed.com
	DOI: 10.4103/tcmj.tcmj_216_20

applied ICG to determine renal blood flow, owing to its fluorescent characteristics. In 1965, Huffman investigated its applicability in detecting cardiac murmurs [5,6]. ICG has since been applied to evaluate physiological brain perfusion [7]. Several perfusion and angiographic applications have been implemented with the advancement of fluorescence imaging.

BIOCHEMICAL AND PHYSICAL PROPERTIES OF INDOCYANINE GREEN

Chemical and optical properties

ICG is a closed-chain, water-soluble cyanine dye with two benzoyl indole moieties [Figure 1]. The molecular weight of ICG is 774.96. Sodium iodide is added to ICG to increase its solubility, which imparts ICG a dark brown color [Figure 2a]. The ICG solution becomes green when diluted with water [Figure 2b]. The excitation wavelength of ICG is 600–900 nm with a maximum wavelength of 875 nm in the blood, whereas the maximum emission wavelength is 830 nm, and emission wavelength range of 750–950 nm [8,9]. The excitation and emission wavelengths are located in the near-infrared (NIR) range, which has the highest tissue penetration depth. ICG is thus an ideal molecular imaging signal source that could be used in various *in vivo* animal models [10].

 $^*Address\ for\ correspondence:$

Dr. Jong-Kai Hsiao,

Department of Medical Imaging, Taipei Tzu Chi Hospital, Buddhist Tzu Chi Medical Foundation, 289, Jianguo Road, Xindian District, New Taipei, Taiwan. E-mail: jongkai@gmail.com

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How to cite this article: Lu CH, Hsiao JK. Indocyanine green: An old drug with novel applications. Tzu Chi Med J 2021; 33(4): 317-22.

Pharmacokinetics

ICG is not absorbed from the gastrointestinal tract. Intravenous injection of ICG is rapidly conjugated by plasma protein (mostly serum albumin) and eliminated by the liver through secretion into the biliary tree in the unconjugated form [11]. The half-life of ICG is 120–240 s [12]. ICG is a high safety drug with an $\rm LD_{50}$ of 80 mg/kg according to an animal study (https://www.drugbank.ca/drugs/DB09374).

Cellular uptake of indocyanine green

The uptake of ICG occurs in the hepatocytes. Various transporters carry ICG into the cytoplasm [13,14]. Organic anion transporting polypeptide1B3 (OATP1B3) and Na+-taurocholate co-transporting polypeptide (NTCP) are two of the principal transporters responsible for the metabolism of ICG in hepatocytes [14]. Transfecting various cancer cell lines using OATP1B3 or NTCP further verified these findings [15-17].

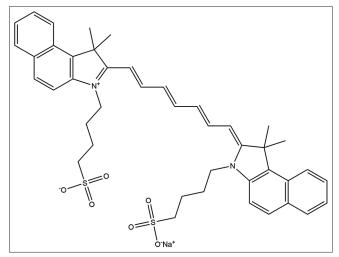


Figure 1: Chemical structure of indocyanine green

Although ICG was transported through transporters similar to bilirubin, findings indicated that intestinal bilirubin transporters are not responsible for the transportation of ICG [18].

CLINICAL PRACTICE OF INDOCYANINE GREEN Retinal evaluation

Fluorescein angiography and ICG angiography are used for retinal evaluation. However, ICG offers a more precise view of choroidal circulation. ICG is a more favorable fluorescent contrast medium than other light-emitting fluorescent dyes because of its infrared fluorescent characteristics. Infrared fluorescent light passes more readily through pigment, turbid fluid, lipid, and hemorrhages, which is common in the optical tract of the eyeball. ICG angiography is favorable for detecting choroidal neovascularization in exudative age-related macular degeneration compared with fluorescein angiography [19,20]. ICG angiography is also an adjunctive tool for differentiating retinal or choroidal tumors[21] and has been applied to aid in diagnosing choroidal inflammatory disorders [22]. Moreover, ICG has been used to detect various pathologic conditions, including vascularized retinal pigment epithelium (RPE) detachment, polypoidal choroidal vasculopathy, traumatic choroidal rupture, diabetic maculopathy, central serous retinopathy, and macular drusen [23]. Although ICG angiography can cause phototoxicity in human RPE cells, which is caused by oxidative damage under continuous ambient light [24], the adverse effects of ICG angiography are very low, and thus ICG is considered a very safe ophthalmic examination [25].

Liver function test

The liver is the largest solid internal organ in the human body. Therefore, major liver surgery remains a high-risk procedure. The hepatic functional reserve of the liver is immediately reduced following injury (e.g., hepatectomy). ICG is an excellent, noninvasive, quantitative, dynamic assessment tool



Figure 2: (a) Indocyanine green in the original constitution with the addition of sodium iodide, displaying a dark brown color. (b) Indocyanine green diluted with phosphate-buffered saline (1 mg/mL) displaying a green color

for evaluating liver function among patients in the intensive care unit or during major liver surgery [26]. The plasma disappearance rate (PDR) and fractional retention at 15 min (R15) are the two parameters used to measure ICG clearance during perioperative liver function monitoring [27].

Most patients requiring liver resection surgery have underlying liver disease. Preoperative assessment of liver function is crucial in selecting candidates for safe hepatic resection to prevent postoperative hepatic failure and death after hepatectomy [28]. Makuuchi et al. reported that ICG R15 plays a determining role in hepatic resection outcomes among patients with small hepatocellular carcinomas [29]. Including ICG R15 as an indicator has improved the operative safety of cirrhotic liver resection [30]. Schwarz et al. determined that ICG is a prognostic factor in both major and minor liver surgeries and proposed that the optimal cutoff value to accurately predict liver dysfunction was PDR <19.5%/min and R15 >5.6% for preoperative ICG clearance test, based on a study in a Western population [31]. ICG R15 is now routinely used for preoperative liver function assessments in numerous countries, including Taiwan [32].

ICG also has various valuable intraoperative uses. Intraoperative measurements of the ICG clearance rate are useful to estimate patients' future liver remnant function during hepatectomy, which helps to determine the preservation of a sufficient amount of liver parenchyma and prevents postoperative liver failure [33]. Furthermore, fluorescence imaging using ICG is promising for intraoperative monitoring during liver surgery [34], and it has been used in liver tumor localization during surgery [35]. ICG fluorescence imaging also helps to identify additional small surface liver tumors during liver resection for cancer [36]. Kobayashi et al. reported five types of ICG fluorescence staining techniques to identify portal vein (PV) territory identification during liver resection, which included transhepatic PV injection and intravenous injection of ICG. These techniques may help to accurately identify the anatomic location during surgery [36]. An increasing number of intraoperative ICG uses during liver surgery are under investigation and require validation in future research studies.

APPLICATIONS UNDER CLINICAL TRIALS Indocyanine green application in general abdominal surgery

The high tissue penetrance, low tissue absorption, and high visibility of critical but small vasculature using ICG have garnered attention in the surgical field [37]. Over 100 clinical trials registered in the USA are currently verifying the feasibility of ICG-assisted imaging guidance for the surgical removal of tumors and metastatic lymph nodes [38-40]. ICG metabolization into the bile duct enables visualization of the bile duct during cholecystectomy. Moreover, bile duct injuries are the most common complication during cholecystectomies. More than 4% of laparoscopic cholecystectomy for acute cholecystitis cause varying degrees of bile duct injury [41]. Consequently, ICG exhibits an operational decisional role by identifying cholecystic ducts during laparoscopic cholecystectomies [42] and can aid in determining the adequate perfusion

for colectomy by intravenous injection of ICG [43]. The proximal transection lines of 112 colorectal cancer patients were evaluated. The intravenous injection of ICG successfully demonstrated that the appropriate transection line prevents anastomotic surgery.

Indocyanine green in fluorescence-guided chest surgery

Video-assisted thoracoscopic surgery (VATS) has become the mainstream thoracic surgery. However, identification of vasculature during operation is sometimes difficult because of the limited field of view. Furthermore, observation of the vasculature may be hindered by other tissue or organs. However, the vasculature could be visualized clearly using ICG injection, even when blocked by lung tissue, in which air space contributes to most of the lung volume. Ligation of the segmental artery and injection of ICG into the peripheral systemic vein during VATS segmentectomy facilitates the identification of segmental lines [44]. Furthermore, ICG has been used to determine the perfusion status of the anastomosis during esophagectomy and gastric tube reconstruction [45]. Lymphatic drainage is difficult to visualize in a standard light source; however, with the assistance of fluorescent thoracoscopy, ICG delivered subcutaneously and eventually drained into the thoracic duct can be identified clearly. This new technique could thus facilitate repair of thoracic duct leakage [46].

An increasing number of trivial occult lung cancers have been diagnosed as the use of computed tomography in early lung cancer detection increases. Certain small tumors are difficult to identify through palpation during surgical removal. ICG has been used to visualize these tumors. ICG is delivered intravenously 24 h before the surgical procedure. Notably, some occult tumors that cannot be recognized by other examinations or palpation were detected after ICG delivery. These occult tumors were later demonstrated to be malignant in most cases [47,48]. The authors also reported that the detection depth was 1.3 cm, and the smallest diameter of the nodule was 0.2 cm [47]. No conclusions regarding the mechanism of ICG accumulation in these malignant tumors have been drawn. However, the enhanced permeability and retention effect is believed to be the most likely underlying mechanism [49].

Sentinel lymph node dissection

Sentinel lymph node dissection is the standard procedure for breast cancer surgery. The identification of sentinel lymph nodes is usually performed using blue dye or radioisotope methods. ICG has been proposed for sentinel lymph node dissection, and clinical trials have been conducted [50,51]. A prospective study reported that up to 89% of sentinel lymph nodes were detected. The authors determined that the limiting factor compared with the radioisotope method, which could reach 96% sensitivity, was a body mass index >40. ICG has also been used for melanoma sentinel lymph node biopsy in 14 cases with the administration of both radioisotope and ICG. The sentinel lymph node detection results are comparable [52]. Despite the lower sensitivity of ICG compared with that of the radioisotope method, ICG is less expensive, has real-time imaging capability, and has no severe adverse effect. Therefore, ICG remains an avenue for sentinel lymph node dissection but requires the development of more sensitive fluorescent detection. The sentinel lymph node detection results are comparable [52].

TRANSLATIONAL AND EXPERIMENTAL INVESTIGATIONS OF INDOCYANINE GREEN Photodynamic therapy

ICG is also a photosensitizer (a compound that can transfer optical energy into reactive oxygen species), which may be used to eradicate tumors. Several *in vitro* and *in vivo* studies on the hepatoma cell line Huh-7 have verified the efficacy of ICG at a dose of 5 mg/kg [53]. The ICG was also encapsulated into nanoparticles, such as liposomes and lactosomes. The photodynamic therapy efficacy of the liposomes and lactosomes was verified on gall bladder cancer cell lines and *in vivo* mouse xenografts [54].

Photothermal therapy

The thermal energy of temperatures above $42.5^{\circ}C$ can destroy tissue, enabling tumor ablation [55]. The photothermal effects of ICG have been investigated in several cell lines, including breast cancer 4T1 cells and animal xenograft models [56]. A study treated 4T1 tumor-bearing mice with NIR LASER at an intensity of 1 W/cm² and ICG at a concentration of 0.1 mg/mL (total quantity of ICG was 200 μ L). Intratumoral injections were administered following LASER ablation. Significant tumor necrosis and lymphocytes infiltrate were observed with histochemical observation.

APPLICATIONS IN BIOMEDICAL RESEARCH Indocyanine green as a reporter probe

ICG is metabolized in the liver where varieties of cell membrane transporters are involved in ICG transportation. These transporters are ideal reporter systems that could be used for *in vivo* molecular imaging such as transporter–ICG combination. The most widely investigated ICG-related reporter protein is OATP1B3, which can carry both ICG and gadoxetate disodium (Gd-EOB-DTPA), a magnetic resonance imaging contrast agent. Research on the feasibility of ICG-OATP1B3 has been conducted, and the results demonstrated that OATP1B3 has dual imaging capability [17]. NTCP is another protein responsible for transporting ICG into the cytoplasm. The efficacy of the ICG-NTCP combination reporter system has also been demonstrated both *in vitro* and *in vivo* [16].

Indocyanine green as an imaging indicator for drug screening

NTCP is the portal for hepatitis B virus (HBV) and hepatitis D virus (HDV) infections into the hepatocyte. Blocking the NTCP with an inhibitor is a prevention strategy for HBV infection. NTCP is a portal for both ICG and HBV into the cytoplasm. High-throughput screening for inhibitors could be performed by identifying the intracellular intensity of ICG, which would indicate the ideal inhibitors for HBV/HDV infection prevention. This concept has been verified in cellular and animal studies [16].

Apical sodium-dependent bile salt transporters (ASBTs), which are responsible for nonalcoholic fatty liver disease,

have also been investigated [57]. ICG is transported by ASBTs and could thus be used as an indicator for *in vitro* drug screening. A conceptual experiment has been established [58].

Photoacoustic imaging using indocyanine green

Photoacoustic imaging is an emerging technology that uses laser targeting on chromophores to emit sound, which is collected for imaging [59]. Endogenous chromophores, such as melanin or ICG, are candidates for photoacoustic imaging. ICG has advantages over other chromophores because of its NIR optical characteristics which are associated with higher tissue depth detection. These conceptual imaging systems have been verified through *in vivo* lung cancer allograft detection in mice. The efficacy of ICG in tumor vascular leakage photoacoustic imaging has been investigated [60].

Conclusion

Because ICG was unveiled in the clinical field, it has displayed superior optical characteristics that have not been surpassed by new technologies. The short half-life of <5 min, high safety dosage, and long-term historical usage of ICG have increased its applicability in numerous new clinical fields, such as photodynamic therapy, photoacoustic imaging, and reporter systems. ICG has the potential to be applied for personalized and precise cancer treatments.

Acknowledgments

We thank I-Jui Hsu for his drawing of the chemical structure of ICG.

Financial support and sponsorship

This research was funded by the Buddhist Tzu Chi Medical Foundation (TCRD-TPE-106-34).

Conflicts of interest

Dr. Jong-Kai Hsiao, an editorial board member at *Tzu Chi Medical Journal*, had no role in the peer review process of or decision to publish this article. The other author declared no conflict of interest in writing this paper.

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