Tzu Chi Medical Journal 24 (2012) 39-42

Contents lists available at SciVerse ScienceDirect

Tzu Chi Medical Journal

journal homepage: www.tzuchimedjnl.com

Review Article

Schizophrenia as a neuronal synaptic disorder related to multiple rare genetic mutations

Yu-Chih Shen^{a,b,*}, Chia-Hsiang Chen^{c,d}

^a Department of Psychiatry, Buddhist Tzu Chi General Hospital, Hualien, Taiwan

^b School of Medicine and Department of Human Development, Tzu Chi University, Hualien, Taiwan

^c Division of Mental Health and Addiction Medicine, Institute of Population Health Sciences, National Health Research Institutes, Miaoli, Taiwan

^d Department of Psychiatry, Chang Gung Memorial Hospital, Linkou and Chang Gung University School of Medicine, Taoyuan, Taiwan

A R T I C L E I N F O

Article history: Received 11 February 2012 Received in revised form 28 February 2012 Accepted 29 February 2012

Keywords: Association and functional studies GAP-43 NRGN Rare variants Schizophrenia SYP

ABSTRACT

Schizophrenia is a highly heritable disorder, but many aspects of its etiology and pathophysiology remain poorly understood. Synaptic pathology has been reported as a feature of the brain in schizophrenia. Abnormal expression of some synaptic proteins (e.g., SYP, GAP-43, and NRGN) in different brain regions has been linked to this disorder in postmortem brain studies. In our series of genetic studies, we used a resequencing strategy to search for genetic variants in these candidate genes in a sample of patients with schizophrenia and nonpsychotic controls, all of whom were Han Chinese from Taiwan, and conducted further association and functional studies. After resequencing these candidate genes, no common polymorphisms appeared to play a major role in conferring susceptibility to schizophrenia in our population. In contrast, we identified some rare patient-specific variants. The results of the reporter gene assays and software analysis demonstrated the influence of reporter genes on the function of each studied gene, suggesting that they may contribute to the pathogenesis of schizophrenia. These data lend support to the hypothesis that multiple rare mutations are involved in the pathogenesis of schizophrenia, and provide genetic clues that indicate the involvement of synaptic pathology in this disorder.

Copyright © 2012, Buddhist Compassion Relief Tzu Chi Foundation. Published by Elsevier Taiwan LLC. All rights reserved.

1. Introduction

Schizophrenia is a complex disorder with a high degree of genetic influence in its etiology [1]. It is now generally accepted that this illness involves variants in multiple genes that are individually insufficient to cause the illness, but which act in combination and with environmental factors to increase the risk of development [2]. Although the majority of the genetic loci that contribute to schizophrenia most likely have weak effects, their identification is essential to determining the neurobiological molecules that play a crucial role in the disorder.

Over the past two decades, structural anomalies have been identified in the brains of patients with schizophrenia [3]. This can be seen in *in vivo* neuroimaging studies that have demonstrated significant ventricular enlargement and decrease in cortical mass [4]. Postmortem brain studies have shown a reduction in the total brain volume, particularly in the cerebral cortex [5], and functional

* Corresponding author. Department of Psychiatry, Buddhist Tzu Chi General Hospital, 707, Section 3, Chung-Yang Road, Hualien, Taiwan. Tel.: +886 3 8565301x2161; fax: +886 3 8466863.

E-mail address: shengmp@so-net.net.tw (Y.-C. Shen).

brain-imaging studies have indicated impaired connectivity between the frontal lobe and other brain regions [6,7]. At the cellular and molecular level, microscopic histopathological studies have demonstrated reduced neuronal size and decreased density of the dendritic spines [8]. Taken together, these changes in the synaptic components may reflect a decrease in the cortical volume, and it is believed that such changes may underlie the aberrant functional connectivity in schizophrenic patients [9,10].

For these reasons, some synaptic proteins have been utilized as proxy markers of synapses to determine whether synaptic alterations are a feature of schizophrenia [9,10]. Three such synaptic proteins that have been repeatedly reported to be involved in schizophrenia are synaptophysin (SYP), growth-associated protein 43 (GAP-43), and neurogranin (NRGN) [9].

2. Genetic and functional analyses of the SYP gene in schizophrenia

SYP is an abundant integral membrane protein in the synaptic vesicles that is expressed in 95% of cortical synaptic terminals [11,12]. Its expression occurs early in neurogenesis and is greatly upregulated during synaptogenesis [13]. This protein is known to

1016-3190/\$ - see front matter Copyright © 2012, Buddhist Compassion Relief Tzu Chi Foundation. Published by Elsevier Taiwan LLC. All rights reserved. doi:10.1016/j.tcmj.2012.03.005





regulate neurotransmitter release and synaptic plasticity [14,15]. In addition, it participates in the biogenesis and recycling of synaptic vesicles [16,17].

Several studies have examined SYP expression in the postmortem brains of patients with schizophrenia. Using *in situ* hybridization, histochemistry, immune autoradiography, and Western blot analysis, decreased SYP expression has been demonstrated in the prefrontal cortex, medial temporal cortex, visual association cortex, hippocampus, and thalamus [18–33]. Conversely, elevated SYP protein levels have been found in the anterior cingulate cortex [34,35]. Microarray studies have also highlighted lower levels of SYP in the postmortem brains of patients with schizophrenia, along with other presynaptic markers [36]. These findings lend support to the notion that SYP disturbance in specific brain regions might be part of the pathogenesis of schizophrenia [9].

The *SYP* gene (gene ID: 6855) has been mapped to chromosome Xp11.23-p11.22. This region has been linked to schizophrenia [37,38]. The function of *SYP*, to a large extent, is believed to be affected by its genetic variations [39]. To test this possibility, we searched for genetic variants in the promoter region, including all exons and both UTR ends of the *SYP* gene, using direct sequencing in a sample of patients with schizophrenia (n = 586) and nonpsychotic controls (n = 576), all of whom were Han Chinese from Taiwan, and conducted further association and functional studies [40].

After sequencing all of the amplicons in the 586 patients and 576 control patients, we identified two common polymorphisms (c.*4+271A>G and c.*4+565T>C) in the SYP gene. Single nucleotide polymorphism (SNP)- and haplotype-based analyses indicated no associations with schizophrenia. In addition, we identified six rare variants in seven of the 586 schizophrenic patients, including one variant (g.-511T>C) located in the promoter region, one synonymous (A104A) variant and two missense variants (G293A and A324T) located in the exonic regions, and two variants (c.*31G>A and c.*1001G>T) located at the 3'UTR. No rare variants were found in the control subjects. The results of the reporter gene assay demonstrate the influence of g.-511T>C and c.*1001G>T on the regulatory function of the SYP gene, while the influence of c.*31G>A may be tolerated. In silico analysis demonstrated the functional relevance of other rare variants. These findings lend support to the hypothesis of multiple rare mutations in schizophrenia and provide genetic clues that indicate the involvement of SYP in this disorder.

3. Genetic and functional analyses of the *GAP*-43 gene in schizophrenia

GAP-43 is a neuron-specific phosphoprotein that is localized to the presynaptic membrane and is a substrate of protein kinase C (PKC). Its phosphorylation by PKC in response to extracellular guidance cues could regulate the behavior of F-actin in neuronal growth cones [41]. In transgenic mice, overexpression of GAP-43 results in the spontaneous formation of new synapses and enhances sprouting after injury [42,43], whereas manipulations that abolish GAP-43 expression result in the disruption of axon outgrowth and could lead to premature death [44,45]. GAP-43 is expressed primarily during brain development and declines sharply in most brain regions after synaptogenesis is completed [41]. High levels of GAP-43 persist in neocortical-association areas and the limbic system throughout life, where the protein might play important roles in mediating experience-dependent synaptic plasticity and long-term potentiation [41].

Several studies have examined GAP-43 expression in the postmortem brains of patients with schizophrenia. Alterations in GAP-43 mRNA levels have been demonstrated in the dorsolateral prefrontal cortex, primary visual cortex, anterior cingulate gyrus, and hippocampus [46–48]. Additionally, GAP-43 protein levels have been found to be altered in the frontal cortex, visual association cortex, and hippocampus [31,49,50]. These findings lend support to the notion that GAP-43 disturbances in specific brain regions might be part of the pathogenesis of schizophrenia.

The *GAP*-43 gene (gene ID: 2596) has been mapped to chromosome 3q13.1-q13.2 [41]. This region has been linked to schizophrenia in a Japanese single multiplex pedigree, and in a meta-analysis of 32 genome-wide linkage studies that were performed on different populations [51,52]. As part of our series on molecular genetic studies on schizophrenia, we were interested in understanding whether the *GAP*-43 gene plays a role in conferring genetic liability to schizophrenia. To test this possibility, we searched for genetic variants in the promoter region and three exons (including both UTR ends) of the *GAP*-43 gene using direct sequencing of a sample of patients with schizophrenia (n = 586) and nonpsychotic controls (n = 576), all of who were Han Chinese from Taiwan, and conducted further association and functional studies [53].

After sequencing all of the amplicons of the 586 patients and 576 control patients, we identified 11 common polymorphisms in the GAP-43 gene. SNP- and haplotype-based analyses indicated no associations with schizophrenia. Additionally, we identified four rare variants in five of the 586 patients, including one variant located in the promoter region (c.-258-4722G>T) and one synonymous (V110V) and two missense (G150R and P188L) variants located on exon 2. No rare variants were found in the control patients. The results of the reporter gene assay demonstrate that the regulatory activities of constructs containing c.-258-4722T was significantly lower when compared with the wild-type construct (c.-258-4722G). In silico analysis also demonstrated the functional relevance of other rare variants. These findings lend support to the hypothesis of multiple rare mutations in schizophrenia, and they provide genetic clues that indicate the involvement of GAP-43 in this disorder.

4. Genetic and functional analyses of the *NRGN* gene in schizophrenia

NRGN is a neural-specific, calmodulin (CaM)-binding protein localized to the postsynaptic membrane and is a substrate of PKC [54]. Glutamate stimulation of N-methyl-D-aspartate (NMDA) receptors results in calcium influx to the neuron and NRGN oxidation [55]. These induce dissociation of the NRGN-CaM complex and stimulate the phosphorylation of NRGN by PKC, which prevents the rebinding of NRGN and CaM [56]. As a CaM reservoir, NRGN regulates the release of CaM and the activities of downstream CaM-Ca²⁺-dependent enzymes that play important roles in the neuroplasticity mechanisms of learning and memory [57,58]. Therefore, altering NRGN activity could mimic the effects of NMDA-receptor hypofunction that has been suggested by several studies, thereby implicating NRGN in the pathophysiology of schizophrenia [59].

NRGN has been found in neurons in the cerebral cortex, hippocampus, striatum, and amygdala [60]. During development, NRGN expression is regulated by thyroid hormones [61], and its highest expression is coincident with the developmental period characterized by rapid dendritic growth and the formation of the majority of the cortical synapses [62]. Broadbelt et al. (2006) examined NRGN expression in the postmortem brains of patients with schizophrenia. Lowered NRGN immunoreactivity was demonstrated in areas 9 and 32 of the schizophrenic prefrontal cortex [63]. Therefore, NRGN disturbance in specific brain regions might be part of the pathogenesis in schizophrenia. Genome-wide association (GWA) studies have identified one SNP (rs12807809) located upstream of the *NRGN* gene that is associated with schizophrenia [64,65], but causal variants that account for the association signal have not been determined. GWA data usually indicate the indirect association of a proxy of a strongly correlated causal variant that has a similar frequency or synthetic association with one or more rarer causal variants in the linkage disequilibrium [66,67]. Causal variants require extensive resequencing and association analysis. In order to find the causal variants of the *NRGN* gene associated with schizophrenia, we searched for genetic variants in the promoter region and all exons (including both UTR ends and rs12807809) using direct sequencing in a sample of patients with schizophrenia (n = 346) and nonpsychotic controls (n = 345), all of whom were Han Chinese from Taiwan, and conducted further association and functional studies [68].

After sequencing all the amplicons of the 346 patients and 345 control subjects, we identified seven common polymorphisms in the NRGN gene. SNP- and haplotype-based analyses indicated no associations with schizophrenia. Additionally, we identified five rare variants in six of the 346 patients, including three rare variants located in the promoter region (g.-620A>G, g.-578C>G, and g.-344G>A) and two rare variants located at 5'UTR (c.-74C>G, and c.-41G>A). No rare variants were found in the control patients. The results of the reporter gene assay demonstrate that the regulatory activities of constructs containing g.-620G, g.-578G, g.-344A, c.-74G, or c.-41A are significantly lower when compared to the wild-type construct. In silico analysis also demonstrated their influence on the regulatory function of the NRGN gene. These data also lend support to the hypothesis that multiple rare mutations are involved in schizophrenia, and provide genetic clues that indicate the involvement of NRGN in this disorder.

5. Summary and future research

In our series of genetic studies of schizophrenia, we used a resequencing strategy to search for genetic variants in each candidate gene in a sample of schizophrenic and control patients, and assessed their associations with schizophrenia. Three candidate genes related to synaptic pathology in schizophrenia were resequenced, but no common polymorphisms appeared to play a major role in conferring susceptibility to schizophrenia in our population. Additionally, we identified some rare patient-specific variants. The results of reporter gene assays and software analysis demonstrate the influence of reporter genes on the function of each studied gene, suggesting that they may contribute to the pathogenesis of schizophrenia.

Our findings support the "common disease, rare alleles" model for explaining some cases of schizophrenia [69]. The hypothesis is that the many mutations that predispose an individual to developing schizophrenia are highly penetrant and individually rare, sometimes even specific to a single patient or family. In this model, different patients harbored different mutations, either in the same gene or in different genes, but each one carried only one or two mutations. Given the fact that these individually rare variants may not contribute, to a significant degree, to the heritability of schizophrenia, their discovery is likely to be much more rewarding than that of common polymorphisms in terms of practical applications, including our understanding schizophrenia's etiology.

After identifying these rare patient-specific variants that are related to synaptic pathology, family studies on the patients with these rare variants should be conducted in order to clarify their inheritance model, genotype/phenotype correlation, phenotypic variability, and penetrance rate. Furthermore, cell-based and electrophysiological experiments are warranted to verify their influence on signal transduction, membrane potential, cell proliferation, migration, and differentiation (e.g., axonal outgrowth, dendrite branching, and synaptogenesis). Furthermore, these findings will be essential to the development of model animals, further pathogenic studies, and novel drugs that could be used to treat this devastating disorder.

It is noteworthy that the targeted resequencing of genes has been used to successfully find associations between rare variants with quantitative traits. However, this approach is currently limited to selected candidate genes. Recently, massive parallel-sequencing technologies, in conjunction with new DNA-enrichment technologies (e.g., exome capture), have been developed that allow the sequencing of targeted regions in large samples of the human genome [70]. In addition, exome capture allows unbiased investigations into complete protein-coding regions of the genome. Because rare variants are usually associated with a high rate of penetrance, they may be much more likely to become the basis for some sort of personalized medicine than those usually discussed in relation to the common polymorphisms.

Acknowledgments

Funding for this study was provided by the National Science Council of Taiwan (grant no. NSC 99-2314-B-303-010-MY3).

References

- Tandon R, Keshavan MS, Nasrallah HA. Schizophrenia, "just the facts": what we know in 2008, II: epidemiology and etiology. Schizophr Res 2008;102: 1–18.
- [2] van Os J, Rutten BP, Poulton R. Gene-environment interactions in schizophrenia: review of epidemiological findings and future directions. Schizophr Bull 2008;34:1066–82.
- [3] Harrison PJ. The neuropathology of schizophrenia. A critical review of the data and their interpretation. Brain 1999;122:593–624.
- [4] Lawrie SM, Abukmeil SS. Brain abnormality in schizophrenia. A systematic and quantitative review of volumetric magnetic resonance imaging studies. Br J Psychiatry 1998;172:110–20.
- [5] Pakkenberg B. Stereological quantitation of human brains from normal and schizophrenic individuals. Acta Neurol Scand Suppl 1992;137:20–33.
- [6] Friston KJ, Frith CD. Schizophrenia: a disconnection syndrome? Clin Neurosci 1995;3:89–97.
- [7] McGuire PK, Frith CD. Disordered functional connectivity in schizophrenia. Psychol Med 1996;26:663-7.
- [8] Selemon LD, Goldman-Rakic PS. The reduced neuropil hypothesis: a circuit based model of schizophrenia. Biol Psychiatry 1999;45:17–25.
- [9] Eastwood SL. The synaptic pathology of schizophrenia: is aberrant neurodevelopment and plasticity to blame? Int Rev Neurobiol 2004;59:47–72.
- [10] Frankle WG, Lerma J, Laruelle M. The synaptic hypothesis of schizophrenia. Neuron 2003;39:205–16.
- [11] Fykse EM, Takei K, Walch-Solimena C, Geppert M, Jahn R, De Camilli P, et al. Relative properties and localizations of synaptic vesicle protein isoforms: the case of the synaptophysins. J Neurosci 1993;13:4997–5007.
- [12] Sudhof TC, Lottspeich F, Greengard P, Mehl E, Jahn R. A synaptic vesicle protein with a novel cytoplasmic domain and four transmembrane regions. Science 1987;238:1142–4.
- [13] Marazzi G, Buckley KM. Accumulation of mRNAs encoding synaptic vesiclespecific proteins precedes neurite extension during early neuronal development. Dev Dyn 1993;197:115–24.
- [14] Alder J, Kanki H, Valtorta F, Greengard P, Poo MM. Overexpression of synaptophysin enhances neurotransmitter secretion at Xenopus neuromuscular synapses. J Neurosci 1995;15:511–9.
- [15] Alder J, Lu B, Valtorta F, Greengard P, Poo MM. Calcium-dependent transmitter secretion reconstituted in Xenopus oocytes: requirement for synaptophysin. Science 1992;257:657–61.
- [16] Daly C, Sugimori M, Moreira JE, Ziff EB, Llinas R. Synaptophysin regulates clathrin-independent endocytosis of synaptic vesicles. Proc Natl Acad Sci USA 2000:97:6120-5.
- [17] Thiele C, Hannah MJ, Fahrenholz F, Huttner WB. Cholesterol binds to synaptophysin and is required for biogenesis of synaptic vesicles. Nat Cell Biol 2000;2:42–9.
- [18] Chambers JS, Thomas D, Saland L, Neve RL, Perrone-Bizzozero NI. Growthassociated protein 43 (GAP-43) and synaptophysin alterations in the dentate gyrus of patients with schizophrenia. Prog Neuropsychopharmacol Biol Psychiatry 2005;29:283–90.
- [19] Davidsson P, Gottfries J, Bogdanovic N, Ekman R, Karlsson I, Gottfries CG, et al. The synaptic-vesicle-specific proteins rab3a and synaptophysin are reduced in

thalamus and related cortical brain regions in schizophrenic brains. Schizophr Res 1999;40:23–9.

- [20] Eastwood SL, Harrison PJ. Synaptic pathology in the anterior cingulate cortex in schizophrenia and mood disorders: a review and a Western blot study of synaptophysin, GAP-43 and the complexins. Brain Res Bull 2001;55:569-78.
- [21] Eastwood SL, Cotter D, Harrison PJ. Cerebellar synaptic protein expression in schizophrenia. Neuroscience 2001;105:219–29.
- [22] Eastwood SL, Harrison PJ. Detection and quantification of hippocampal synaptophysin messenger RNA in schizophrenia using autoclaved, formalin-fixed, paraffin wax-embedded sections. Neuroscience 1999;93:99–106.
- [23] Eastwood SL, Burnet PW, Harrison PJ. Altered synaptophysin expression as a marker of synaptic pathology in schizophrenia. Neuroscience 1995;66: 309–19.
- [24] Eastwood SL, Harrison PJ. Decreased synaptophysin in the medial temporal lobe in schizophrenia demonstrated using immunoautoradiography. Neuroscience 1995;69:339–43.
- [25] Glantz LA, Lewis DA. Reduction of synaptophysin immunoreactivity in the prefrontal cortex of subjects with schizophrenia. Regional and diagnostic specificity. Arch Gen Psychiatry 1997;54:943–52.
- [26] Halim ND, Weickert CS, McClintock BW, Hyde TM, Weinberger DR, Kleinman JE, et al. Presynaptic proteins in the prefrontal cortex of patients with schizophrenia and rats with abnormal prefrontal development. Mol Psychiatry 2003;8:797–810.
- [27] Hemby SE, Ginsberg SD, Brunk B, Arnold SE, Trojanowski JQ, Eberwine JH. Gene expression profile for schizophrenia: discrete neuron transcription patterns in the entorhinal cortex. Arch Gen Psychiatry 2002;59:631–40.
- [28] Karson CN, Mrak RE, Schluterman KO, Sturner WQ, Sheng JG, Griffin WS. Alterations in synaptic proteins and their encoding mRNAs in prefrontal cortex in schizophrenia: a possible neurochemical basis for 'hypofrontality'. Mol Psychiatry 1999;4:39–45.
- [29] Landen M, Davidsson P, Gottfries CG, Grenfeldt B, Stridsberg M, Blennow K. Reduction of the small synaptic vesicle protein synaptophysin but not the large dense core chromogranins in the left thalamus of subjects with schizophrenia. Biol Psychiatry 1999;46:1698–702.
- [30] Mukaetova-Ladinska EB, Hurt J, Honer WG, Harrington CR, Wischik CM. Loss of synaptic but not cytoskeletal proteins in the cerebellum of chronic schizophrenics. Neurosci Lett 2002;317:161–5.
- [31] Perrone-Bizzozero NI, Sower AC, Bird ED, Benowitz LI, Ivins KJ, Neve RL. Levels of the growth-associated protein GAP-43 are selectively increased in association cortices in schizophrenia. Proc Natl Acad Sci USA 1996;93:14182–7.
- [32] Vawter MP, Howard AL, Hyde TM, Kleinman JE, Freed WJ. Alterations of hippocampal secreted N-CAM in bipolar disorder and synaptophysin in schizophrenia. Mol Psychiatry 1999;4:467–75.
- [33] Webster MJ, Shannon Weickert C, Herman MM, Hyde TM, Kleinman JE. Synaptophysin and GAP-43 mRNA levels in the hippocampus of subjects with schizophrenia. Schizophr Res 2001;49:89–98.
- [34] Gabriel SM, Haroutunian V, Powchik P, Honer WG, Davidson M, Davies P, et al. Increased concentrations of presynaptic proteins in the cingulate cortex of subjects with schizophrenia. Arch Gen Psychiatry 1997;54:559–66.
- [35] Honer WG, Falkai P, Chen C, Arango V, Mann JJ, Dwork AJ. Synaptic and plasticity-associated proteins in anterior frontal cortex in severe mental illness. Neuroscience 1999;91:1247–55.
- [36] Mirnics K, Levitt P, Lewis DA. Critical appraisal of DNA microarrays in psychiatric genomics. Biol Psychiatry 2006;60:163–76.
- [37] Dann J, DeLisi LE, Devoto M, Laval S, Nancarrow DJ, Shields G, et al. A linkage study of schizophrenia to markers within Xp11 near the MAOB gene. Psychiatry Res 1997;70:131–43.
- [38] Wei J, Hemmings GP. Searching for a locus for schizophrenia within chromosome Xp11. Am J Med Genet 2000;96:4–7.
- [39] Wei J, Hemmings GP. A further study of a possible locus for schizophrenia on the X chromosome. Biochem Biophys Res Commun 2006;344:1241–5.
- [40] Shen YC, Tsai HM, Ruan JW, Liao YC, Chen SF, Chen CH. Genetic and functional analysis of the gene encoding synaptophysin in schizophrenia. Schizophr Res 2012. doi:10.1016/j.schres.2012.01.028.
- [41] Benowitz LI, Routenberg A. GAP-43: an intrinsic determinant of neuronal development and plasticity. Trends Neurosci 1997;20:84–91.
- [42] Aigner L, Arber S, Kapfhammer JP, Laux T, Schneider C, Botteri F, et al. Overexpression of the neural growth-associated protein GAP-43 induces nerve sprouting in the adult nervous system of transgenic mice. Cell 1995;83:269–78.
- [43] Holtmaat AJ, Dijkhuizen PA, Oestreicher AB, Romijn HJ, Van der Lugt NM, Berns A, et al. Directed expression of the growth-associated protein B-50/ GAP-43 to olfactory neurons in transgenic mice results in changes in axon morphology and extraglomerular fiber growth. J Neurosci 1995;15:7953–65.
- [44] Routtenberg A. Knockout mouse fault lines. Nature 1995;374:314-5.

- [45] Strittmatter SM, Fankhauser C, Huang PL, Mashimo H, Fishman MC. Neuronal pathfinding is abnormal in mice lacking the neuronal growth cone protein GAP-43. Cell 1995;80:445–52.
- [46] Eastwood SL, Harrison PJ. Hippocampal and cortical growth-associated protein-43 messenger RNA in schizophrenia. Neuroscience 1998;86:437–48.
- [47] Hakak Y, Walker JR, Li C, Wong WH, Davis KL, Buxbaum JD, et al. Genomewide expression analysis reveals dysregulation of myelination-related genes in chronic schizophrenia. Proc Natl Acad Sci USA 2001;98:4746–51.
- [48] Weickert CS, Webster MJ, Hyde TM, Herman MM, Bachus SE, Bali G, et al. Reduced GAP-43 mRNA in dorsolateral prefrontal cortex of patients with schizophrenia. Cereb Cortex 2001;11:136–47.
- [49] Blennow K, Bogdanovic N, Gottfries CG, Davidsson P. The growth-associated protein GAP-43 is increased in the hippocampus and in the gyrus cinguli in schizophrenia. J Mol Neurosci 1999;13:101–9.
- [50] Sower AC, Bird ED, Perrone-Bizzozero NI. Increased levels of GAP-43 protein in schizophrenic brain tissues demonstrated by a novel immunodetection method. Mol Chem Neuropathol 1995;24:1–11.
- [51] Kaneko N, Muratake T, Kuwabara H, Kurosaki T, Takei M, Ohtsuki T, et al. Autosomal linkage analysis of a Japanese single multiplex schizophrenia pedigree reveals two candidate loci on chromosomes 4q and 3q. Am J Med Genet B Neuropsychiatr Genet 2007;144B:735–42.
- [52] Ng MY, Levinson DF, Faraone SV, Suarez BK, DeLisi LE, Arinami T, et al. Metaanalysis of 32 genome-wide linkage studies of schizophrenia. Mol Psychiatry 2009;14:774–85.
- [53] Shen YC, Tsai HM, Cheng MC, Hsu SH, Chen SF, Chen CH. Genetic and functional analysis of the gene encoding GAP-43 in schizophrenia. Schizophr Res 2012;134:239–45.
- [54] Ran X, Miao HH, Sheu FS, Yang D. Structural and dynamic characterization of a neuron-specific protein kinase C substrate, neurogranin. Biochemistry 2003; 42:5143–50.
- [55] Li J, Pak JH, Huang FL, Huang KP. N-methyl-D-aspartate induces neurogranin/ RC3 oxidation in rat brain slices. J Biol Chem 1999;274:1294–300.
- [56] Rodriguez-Sanchez P, Tejero-Diez P, Diez-Guerra FJ. Glutamate stimulates neurogranin phosphorylation in cultured rat hippocampal neurons. Neurosci Lett 1997;221:137–40.
- [57] Huang KP, Huang FL, Jager T, Li J, Reymann KG, Balschun D. Neurogranin/RC3 enhances long-term potentiation and learning by promoting calciummediated signaling. J Neurosci 2004;24:10660–9.
- [58] Pak JH, Huang FL, Li J, Balschun D, Reymann KG, Chiang C, et al. Involvement of neurogranin in the modulation of calcium/calmodulin-dependent protein kinase II, synaptic plasticity, and spatial learning: a study with knockout mice. Proc Natl Acad Sci USA 2000;97:11232–7.
- [59] Tsai G, Coyle JT. Glutamatergic mechanisms in schizophrenia. Annu Rev Pharmacol Toxicol 2002;42:165–79.
- [60] Represa A, Deloulme JC, Sensenbrenner M, Ben-Ari Y, Baudier J. Neurogranin: immunocytochemical localization of a brain-specific protein kinase C substrate. J Neurosci 1990;10:3782–92.
- [61] Dowling AL, Zoeller RT. Thyroid hormone of maternal origin regulates the expression of RC3/neurogranin mRNA in the fetal rat brain. Brain Res Mol Brain Res 2000;82:126–32.
- [62] Iniguez MA, Rodriguez-Pena A, Ibarrola N, Aguilera M, Munoz A, Bernal J. Thyroid hormone regulation of RC3, a brain-specific gene encoding a protein kinase-C substrate. Endocrinology 1993;133:467–73.
- [63] Broadbelt K, Ramprasaud A, Jones LB. Evidence of altered neurogranin immunoreactivity in areas 9 and 32 of schizophrenic prefrontal cortex. Schizophr Res 2006;87:6–14.
- [64] Stefansson H, Ophoff RA, Steinberg S, Andreassen OA, Cichon S, Rujescu D, et al. Common variants conferring risk of schizophrenia. Nature 2009;460: 744–7.
- [65] Ripke S, Sanders AR, Kendler KS, Levinson DF, Sklar P, Holmans PA, et al. Genome-wide association study identifies five new schizophrenia loci. Nat Genet 2011;43:969–76.
- [66] Dickson SP, Wang K, Krantz I, Hakonarson H, Goldstein DB. Rare variants create synthetic genome-wide associations. PLoS Biol 2010;8:e1000294.
- [67] Takeuchi F, Kobayashi S, Ogihara T, Fujioka A, Kato N. Detection of common single nucleotide polymorphisms synthesizing quantitative trait association of rarer causal variants. Genome Res 2011;21:1122–30.
- [68] Shen YC, Tsai HM, Cheng MC, Hsu SH, Chen SF, Chen CH. Genetic and functional analysis of the gene encoding neurogranin in schizophrenia. Schizophr Res 2012. doi:10.1016/j.schres.2012.01.011.
- [69] McClellan JM, Susser E, King MC. Schizophrenia: a common disease caused by multiple rare alleles. Br J Psychiatry 2007;190:194–9.
- [70] Teer JK, Mullikin JC. Exome sequencing: the sweet spot before whole genomes. Human Molecular Genetics 2010;19:R145–51.